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Species Distribution Modelling: Contrasting presence-only models with plot abundance data

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Species distribution models (SDMs) are widely used in ecology and conservation. Presence-only SDMs such as MaxEnt frequently use natural history collections (NHCs) as occurrence data, given their huge numbers and accessibility. NHCs are often spatially biased which may generate inaccuracies in SDMs. Here, we test how the distribution of NHCs and MaxEnt predictions relates to a spatial abundance model, based on a large plot dataset for Amazonian tree species, using inverse distance weighting (IDW). We also propose a new pipeline to deal with inconsistencies in NHCs and to limit the area of occupancy of the species. We found a significant but weak positive relationship between the distribution of NHCs and IDW for 66% of the species. The relationship between SDMs and IDW was also significant but weakly positive for 95% of the species, and sensitivity for both analyses was high. Furthermore, the pipeline removed half of the NHCs records. Presence-only SDM applications should consider this limitation, especially for large biodiversity assessments projects, when they are automatically generated without subsequent checking. Our pipeline provides a conservative estimate of a species' area of occupancy, within an area slightly larger than its extent of occurrence, compatible to e.g. IUCN red list assessments.

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Species distribution models (SDMs) are widely used in the fields of macroecology, biogeography and biodiversity research for modelling species geographic distributions based on correlations between known occurrence records and the environmental conditions at occurrence localities^{1,2}. SDMs generate geographical maps of a species' environmental suitability, its likelihood of being collected, and its local abundance³. Their application includes selecting conservation areas, predicting the effects of climate change on species ranges and determining the risk of species invasions^{4,5}. The wide use of SDMs in ecological and conservation research can partly be explained by the growing availability of georeferenced species records (e.g. GBIF, SpeciesLink) and environmental data (e.g. WorldClim, CliMond)^{6,7} on the web, together with the user-friendly character of some of the modelling methods.

One of the most commonly used SDMs is MaxEnt, which has become increasingly popular since its introduction⁸. This machine-learning algorithm estimates a species' probability distribution that has maximum entropy (closest to uniform), subject to a set of constraints based upon our knowledge of the environmental conditions at known occurrence sites¹. MaxEnt is a presence-only model, enabling scientists to utilize the abundant data sources of natural history collections (NHCs), avoiding the high costs of sampling the species throughout their extent of occurrence. Presence data are abundant, but absence data are hard to obtain and often unreliable due to insufficient survey effort. To counter the lack of absences, MaxEnt uses a background sample to contrast the distribution of presences along environmental gradients against the distribution background points, randomly drawing from the study area.

NHCs, however, may not be independently drawn from the investigated populations due to the non-random nature of collecting^{9,10}. Because collectors aim to collect as many species as possible, rare species are often over-represented in herbaria, whereas common species are underrepresented, producing collectors' bias¹¹. Therefore, the relative number of specimens per species in herbaria is not a good representation of the species' relative abundance in the field. Additionally, NHCs have spatial bias due to geographical differences in survey effort, data storage and mobilization^{9,10,12}. This may have negative impacts on the performance of presence-only SDMs if this results in environmentally biased sampling^{12–15}. Negative impact of spatial bias is not always present, however^{16,17}.

MaxEnt has shown to outperform other SDMs in several studies^{18–22}. Nevertheless, some drawbacks have been identified. For example, MaxEnt may underestimate the probability of occurrence within areas of observed presence, while overestimating it in areas beyond the species' known extent of occurrence²³. Like other SDMs, one essential assumption of MaxEnt is that the presence-data are an independent sample from the species' unknown probability distribution of occurrence over the study area¹. Given the shortcomings of NHCs due to collectors' bias mentioned above, this assumption may not be met.

With a large set of plots with quantitative data, species abundances may be estimated by a spatial interpolation of local species' abundances²⁴. Based on plot data, where all species are collected (regardless of commonness or rarity), the interpolation method arguably suffers less from the collectors' bias and is exclusively based on location. The abundance maps may serve as the species' estimated probability distribution and a higher local abundance implies a higher probability of collecting. That is, the chance of encountering a species is higher in a region where the relative abundance of that species is high, than where the relative abundance is low. With spatially interpolated abundances we may thus test whether NHCs can actually be considered a random sample of the unknown probability distribution.

Here we test how the geographic distribution of NHCs relates to the species relative abundance. To achieve this we address the following questions: (1) Do NHCs represent an independently drawn sample from the unknown probability distribution of a species? And (2) how does MaxEnt's predicted environmental suitability

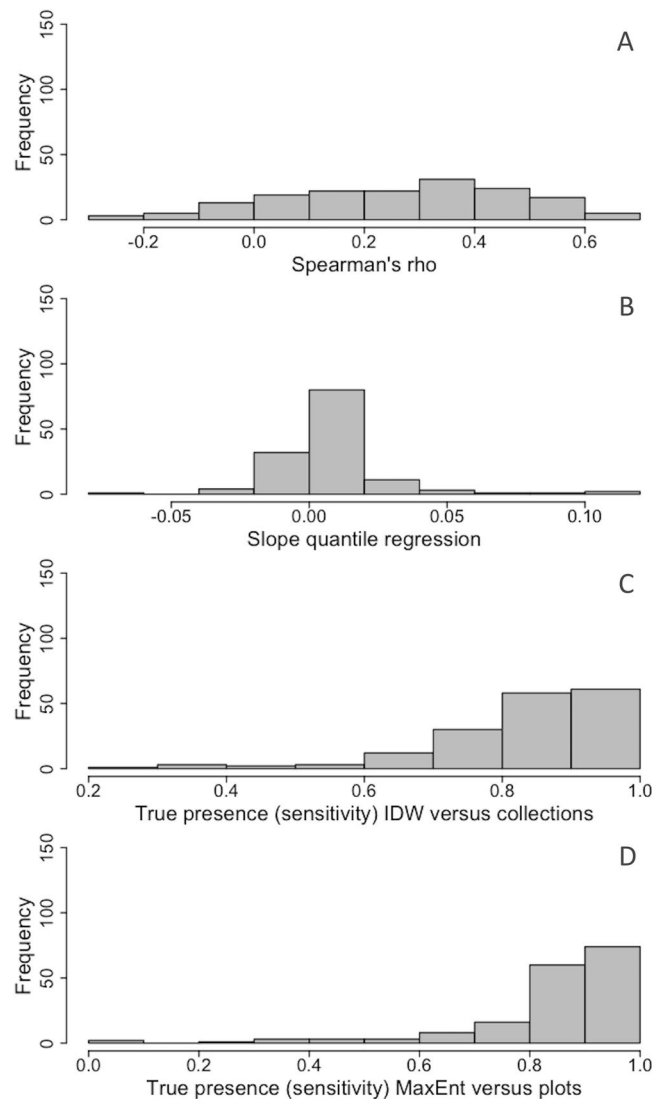


Figure 1. Frequency distributions for 189 significant hyperdominant Amazonian tree species of (A) the Spearman's correlation index ρ between MaxEnt's predicted environmental suitability and relative local abundance of the plots; (B) The slopes of the linear 90th percentile quantile regression between MaxEnt's predicted environmental suitability and the relative local abundance of the plots; (C) The true presence (sensitivity) of the distribution predicted by the IDW maps compared to the collection localities; and (D) The true presence (sensitivity) of the distribution predicted by the MaxEnt maps compared to the plot presence.

compare to plot abundance data and spatial interpolation of species abundances? To answer these questions, we used NHCs and abundance plot data of 227 hyperdominant Amazonian tree species, which are the most common tree species that together make up half of all trees with a diameter (dbh) over 10 cm in Amazonia^{24,25}, the most biodiverse rainforest on Earth. We used NHCs and MaxEnt to construct presence-only SDMs for all 227 species and constructed the abundance maps by spatial interpolation of the plot abundance data for all species as well. To answer the first question, we compared the collection records to the interpolated abundance maps for each species. Secondly, we compared MaxEnt's predicted environmental suitability maps to the same interpolated abundance maps for each of the 227 species.

Results

NHCs data distribution and relative abundance analysis. The analysis testing our first question, whether NHCs are an independent draw from the unknown probability distribution, resulted in a significant ($P < 0.05$), but very weak positive relationship for 149 (66%) species of the 227. For these species the chance of being collected indeed increased slightly with higher interpolated relative abundance. For the other 78 species (34%), this relationship was non-significant or negative (Appendix S1).

Predicted environmental suitability compared to species relative abundances. Further analyses were carried out using only 170 species. Species, that had MaxEnt's predicted environmental suitability not

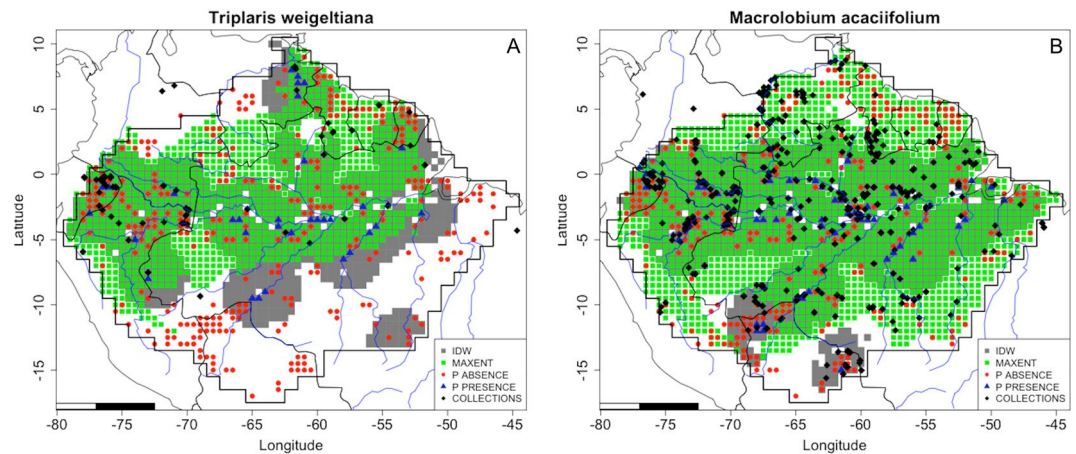


Figure 2. The predicted area of occupancy by MaxEnt (green) and the IDW map (grey) of (A) *Triplaris weigeltiana*; and (B) *Macrolobium acaciifolium*. The localities of the collections, presence and absence plots are also indicated. Maps created with custom R script. Base map source (country.shp, rivers.shp): ESRI (<http://www.esri.com/data/basemaps>, © Esri, DeLorme Publishing Company).

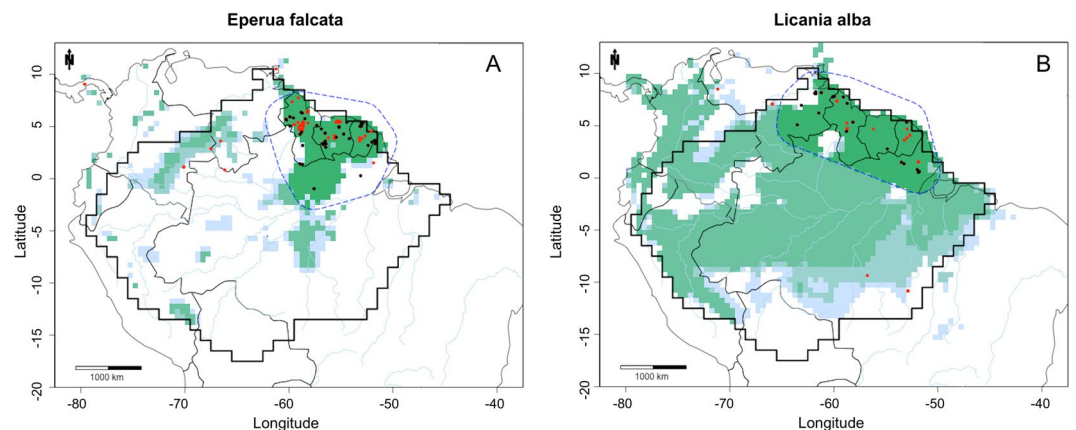


Figure 3. MaxEnt environmental suitability maps for (A) *Eperua falcata*; (B) *Licania alba*. MaxEnt maps constructed using GBIF records, cleaned GBIF records, kernel-density estimate GBIF records, and kernel-density estimate GBIF records plus the buffer clip. **Black dots:** GBIF records. **Red dots:** GBIF records after the use of the cleaning pipeline. **Dashed blue line:** buffer based on a convex hull around species cleaned collections. **Light blue:** predicted environmental suitability using GBIF records. **Light green:** predicted environmental suitability using cleaned GBIF records. **Medium green:** predicted environmental suitability using kernel density estimate GBIF records. **Dark green:** predicted environmental suitability using kernel density estimate GBIF records and the buffer clip, resulting in the final predicted area of occupancy. Maps created with custom R script. Base map source (country.shp, rivers.shp): ESRI (<http://www.esri.com/data/basemaps>, © Esri, DeLorme Publishing Company).

significantly different from a random expectation tested with bias corrected null models, were excluded (57 species). For 161 of the 170 species (95%), MaxEnt's predicted environmental suitability was also significantly correlated with interpolated abundance ($P < 0.05$). The correlations and, thus the biological significance, were low however, with a mean rho (Spearman rank correlation) of 0.26 (Fig. 1A). A linear 90th quantile regression revealed that for 135 (79%) of the 175 species, the logistic output of MaxEnt could significantly ($P < 0.05$) predict the highest 10% of the local relative abundance values. The slope of the regression and thus the biological significance was very low, with a mean slope of only 0.01 (Fig. 1B).

We also investigated the performance of the IDW output against NHCs data and the MaxEnt output against plot presence (sensitivity), to check whether the maps were accurate references to the occurrence data of each other (Appendix S2). Approximately 87% of the grid cells with species' NHCs were correctly predicted as present by the IDW maps with a median true positive rate of 0.87 (Fig. 1C). The same analyses for MaxEnt showed that 88% of the grid cells with plot presence were correctly predicted by MaxEnt maps, with a median true positive rate of 0.88 (Fig. 1D). Sensitivity for both analyses was high.

We provide maps (combined MaxEnt and IDW maps [as in Fig. 2]) for all species in the Supplementary Material S3. The predicted environmentally suitable region and the abundance distribution were similar for very abundant species with a large extent of occurrence, such as *Brosimum rubescens* (Fig. S3_14A), *Conceveiba guianensis* (Fig. S3_32A) and *Eschweilera coriacea* (Fig. S3_49A). The same was true in the case of the species *Clathrotropis glaucophylla* (Fig. S3_30A) and *Cenostigma tocantinum* (Fig. S3_26A), despite the fact that neither species has a wide extent of occurrence.

Moreover, MaxEnt also correctly predicted the environmental unsuitability of non-forested savanna areas, which are located in the north (Brazil, Guianas and Venezuela) and south of the map (northern Bolivia). These close matches apply to very abundant species with a large extent of occurrence, such as *Licania micrantha* (Fig. S3_87A), and *Ocotea aciphylla* (Fig. S3_111A).

For *Triplaris weigeltiana*, a species with a northern Amazonian distribution, MaxEnt also correctly predicted its absence in these northern non-forested areas (Fig. 2A, S3_160A,B). In this case MaxEnt was able to establish a relationship between species distribution and vegetation type, based on climate variables (temperature and precipitation) and species occurrence. For *Macrolobium acaciifolium*, a riverine species, the IDW presented limitations. This species is rarely recorded in plots, because the plots are mostly far from river edges. Thus, the species was found only in plots near to major rivers such as the Amazon. In this case NHCs provided better information about species occurrence, as collectors can reach areas closer to other smaller rivers aiming to collect more species. In such a case, MaxEnt maps presented a wider distribution for the species (Fig. 2B, S3_92A–C).

IDW maps predicted widespread distributions for palms, for which the MaxEnt estimates were in sharp disagreement. Palms species are more difficult to collect, which can result in a lack of specimens in NHCs²⁴. IDW maps appear to be more accurate for these species, because all species are recorded inside plots. In eastern Amazonia this was particularly severe because NHCs showed a large lack of occurrence in comparison with plot data but also proper locations were rejected by a kernel density estimate (KDE), because of the huge amount of palm occurrence data from the Aarhus University Palm Transect Database in western Amazonia. Some of the species affected were *Attalea butyracea* (Fig. S3_8A), *Euterpe precatoria* (Fig. S3_60A), *Iriartea deltoidea* (Fig. S3_72A), *Oenocarpus bacaba* (Fig. S3_113A), *Oenocarpus bataua* (Fig. S3_114A) and *Socratea exorrhiza* (Fig. S3_150A).

NHCs data cleaning treatment and MaxEnt map building. All 227 hyperdominant species had records excluded by the data cleaning treatment, the consequence of records that either lack geographic information, are duplicates at the used grid cell resolution of 0.5 degree or were outliers based on a kernel-density estimate (Appendix S4). An average of 50% of the records was excluded. The first twelve species with the most excluded records were palms, with a mean of 96% of excluded records. The total average of excluded records decreased to 43% when palms were taken out of the analyses (Appendix S4). This high percentage is due to the huge amount of palm occurrence data of the Aarhus University Palm Transect Database²⁶. At this moment this database contains 543,000 records, all available in GBIF. Most of these records represent observations in many plots inside the same grid cell, thus these records were removed and considered as a single observation.

After the kernel density estimate treatment the average of excluded records was 57%, presenting an increment of 6.7% in the total amount of records excluded. *Eperua purpurea* and *Eperua leucantha* collections were in good agreement with plot data distribution, after outliers were excluded by the kernel density estimate (Appendix S5). In the case of *Eperua falcata*, some occurrences in Colombia and Venezuela were in fact misidentifications of *Eperua leucantha*, since this species occurs only in the Guianas (H. ter Steege, pers. obs.). The kernel density estimate function correctly removed these occurrences outside the *E. falcata* cluster observed in the Guianas (Fig. 3A). Some occurrences of *Licania alba* in southeast Amazonia, an area with no plot data, were also removed by the kernel-density estimate function (Fig. 3B).

Because of the use of the buffer treatment to limit MaxEnt predictions around the species' extent of occurrence, MaxEnt maps predicted an area of occupancy close to that of the IDW maps. The median value for MaxEnt's area of occupancy was 1354 0.5-degree grid cells, and the median for IDW was 1217. For 98 (58%) of the 170 species, MaxEnt predicted an area of occupancy bigger than the that predicted by IDW, and for 115 (42%) of the species IDW had a predicted area of occupancy bigger than MaxEnt. In 15% of the cases (26 species) the size difference in area of occupancy was smaller than 5% (Appendix S2).

Discussion

Using NHCs for presence-only SDMs. Collection density was weakly related to relative abundance in most tree species, and for 34% there was no positive relationship between the chance of being collected and local abundance, violating the assumption of MaxEnt that collection localities are an independently drawn sample from a species' unknown probability distribution¹. The differences between the distribution of NHCs and local abundance could limit the ability of presence-only SDMs to predict species probability of distribution as predicted by spatial interpolation of local species' abundance.

MaxEnt's premise that species occurrences are drawn randomly from the unknown probability distribution¹ may not be met for two reasons: (1) collections are spatially biased with regard to environmental conditions¹³; (2) and collections are spatially biased with regard to areas of high abundance, with underrepresentation of in areas of high abundance and overrepresentation in areas of low abundance. Much attention has been given to the possible impacts of spatial bias on the performance of presence-only SDMs, with some showing a negative impact on these models^{13–15}, and others arguing for the robustness of MaxEnt against spatial bias^{17,27}. However, little attention has been given to the second issue. With our plot dataset, we addressed the relationship between collection localities and the predicted spatial abundance distribution.

In 66% of the cases we found that a higher local relative abundance indeed increased the chance of being collected, although the correlations were very weak (Fig. 1A), and the majority of collections originated from areas

with a low relative abundance due to the large areas where a given species' abundance is low. Even hyperdominant species are usually only dominant in one or two of the six regions of Amazonia, most hyperdominant species have a large geographic extent of occurrence but are habitat specialists²⁴. Steege *et al.*²⁵ also found that abundance is a poor predictor for the number of collections of a species compared to the size of its extent of occurrence. Additionally, herbaria are characterized by the earlier discussed collectors' bias¹¹. Although we addressed the spatial bias of survey effort by including a bias based background file in our MaxEnt modelling, the lack of a significant positive relationship between relative abundance estimated by IDW and collection density for many species suggests that this assumption of MaxEnt is not met because of the way species are collected.

MaxEnt maps vs. IDW maps. We also asked if MaxEnt maps would be a close match to the IDW maps. In general, environmental suitability does not reflect a species abundance. Presence-only SDMs, such as MaxEnt, are based on correlations between species presence and environmental conditions, predicting the environmental suitability for a species, and not their realized distribution⁵. Relative abundance in the other hand is based solely on abundance, estimating the number of trees belonging to each species in the grid cells²⁸. The Spearman's rank correlation and the linear 90th percentile quantile regression showed a very weak positive relationship between MaxEnt's predicted environmental suitability and IDW relative abundance prediction at plot localities, contrary to the results of VanDerWal *et al.*²⁹, who found a strong relationship between the two. Their research differs in that they modelled a biogeographical region with tropical and subtropical rainforests, and also drier and warmer environments. The relationship between environmental suitability and local abundance is likely to be stronger when more (extreme) divergent conditions are included, such as areas from different biomes. Perhaps Amazonia's less divergent conditions, representing perhaps a one single biome, in a much larger area, are potentially responsible for this weak relationship.

To test their further predictive performances we also converted both outputs to binary maps. Some studies have addressed questions about the transformation of SDM predictions into discrete representations such as binary maps, aiming to estimate area of occupancy, species richness and others applications^{30–33}. Binary maps can add more uncertainties to model predictions, especially because it is necessary to set a threshold to distinguish between species presence and absence, which can be selected arbitrarily or without taking into account the context of the study. However, we avoided thresholds based on specificity (prediction of absences) because of the lack of absence data³⁴. In many cases our MaxEnt binary maps presented an area of occupancy close to those made with IDW, presenting a high median sensitivity (88%). Moreover our MaxEnt binary maps also correctly predicted absence in naturally non-forested areas in northern Amazonia for many species (Appendix S3).

MaxEnt's environmental suitability mostly predicted much larger area of occupancy than those predicted with the IDW relative abundance. We reduced this effect estimating species extent of occurrence using a convex hull around each species records, plus a buffer of 300 km. This approach minimized MaxEnt's overestimation of the area of occupancy beyond the species' known geographical range (extent of occurrence), over climatically suitable areas, by restricting the species' predicted suitable habitat, providing a more conservative estimate for the species' area of occupancy (Appendix S5)^{35,36}.

The IDW relative abundance models showed an opposite behaviour, underpredicting areas where collections are present but where no plots have recorded the species. The high sensitivity of the MaxEnt compared to those of the IDW is in agreement with a previous study³⁷, where models fit to presence-only data yielded higher sensitivity but a lower specificity than presence-absence models. Nevertheless, in our case, the IDW relative abundance yielded sensitivity rates based on collection localities that were as high as the sensitivity rates of MaxEnt's predicted environmental suitability based on plot presence localities (87%). Thus, both models function similarly in predicting species presences.

Collections versus abundance plot data. In some cases, collections were located outside the species' extent of occurrence predicted by the IDW maps. This divergence follows from the methodical differences between collections and plot assessments. The distributions as predicted by the IDW do not always cover the whole species' extent of occurrence. Because there are only 550 individuals (on average) in one plot, and 16,000 tree species in Amazonia²⁴, one plot obviously cannot contain all species that are present in the surrounding area. Furthermore, many plots, lacking a given species, are within the extent of occurrence predicted by IDW, and many plots with absences are located in close proximity to plots with presence data. This results in low specificity values. NHCs comprise a species' range including areas of low abundance; while plot data have information on abundance, but may miss areas of low abundance, and, thus, may miss rare species more easily.

Environmental suitability versus dispersal limitation. The second large difference between the two models is the theoretical principles they are based upon. MaxEnt is based on environmental suitability, which is appropriate since correlations between species' distributions and climate are evident^{5,36}. Nevertheless, predicting actual (realized) distributions also requires information on biotic interactions, dispersal limitation, and other environmental variables, which are beyond presence-only SDM⁵. IDW, on the other hand, is based on location only. Thus, both models cover only one of the three explanatory variables for species distributions. Again, it will depend on the aim of the research which type of model is most suitable. In either case predicted species distributions need to be interpreted with caution.

Collection data and cleaning pipeline. We propose a cleaning pipeline to remove possible inconsistencies in collection data. Unlike species-specific approaches, many studies use large numbers of species, lacking correction because of the great number of references and specialists to be consulted³⁸. Collection data available in global datacentres, such as GBIF, cannot carry out thorough data-correction procedures, and the quality of the records has been debated and tested in some cases³⁹. Some records have no locality information, or

coordinates are based on cities close to the observed distribution, and may contain duplicated data or zeros as information^{38–40}.

We used a pipeline that cleans collection data by removing records with a lack of geographic information³⁸, and we strongly recommend the use of analytical tools to correct inconsistencies present in global databases. The cleaning process also removed coordinates considered spatial outliers by a kernel-density estimate, omitting locations too far from the central part of the distribution, which we assume to be misidentifications.

Our results suggest that half of the species records are likely inconsistent, missing geographical information, such as latitude, longitude or locality. Palms were the most impacted species, because the huge amount of records available with high levels of redundancy.

We used a kernel density estimate (KDE) to remove geographical outliers of the NHCs. This function removed e.g. occurrences outside the *Eperua falcata* cluster observed in the Guianas (Fig. 3A), and *Licania alba* in south-east Amazonia (Fig. 3B). Although the KDE excluded only a small number of records compared to the previous cleaning step, it was able to identify some isolated occurrences, which we considered likely misidentifications. The KDE, however, showed limitations with palm species, removing some eastern Amazonia records, simply caused by the great number of collections in the Aarhus University Palm Transect Database in western Amazonia.

Conclusion

We have shown that the NHCs violate the assumption of MaxEnt that collection localities are an independently drawn sample from a species' unknown probability distribution. Although we found a relationship between NHCs and relative abundance for some species, it was very weak. Additionally, we found that the majority of MaxEnt's predicted environmental suitability values differ from those of the IDW relative abundance values, and its results cannot be interpreted as an abundance estimate. Nevertheless, MaxEnt predicts probability of occurrence well, and both models largely overlap and predict similar areas of occupancy, showing high sensitivities. Furthermore, NHCs data should undergo cleaning processes before being used to represent occurrences in species distribution models. We showed that, half of the species records are likely inconsistent, missing geographical information, such as latitude, longitude or locality, and it also may represent misidentifications of the species. We therefore conclude that distribution maps as generated by MaxEnt should be used with caution. Their application should not be based solely on unsupervised models, especially because their easily constructed distribution maps are tempting to utilize without indication of probable errors. This outcome is particularly important for biodiversity assessments, for which SDMs of a large number of species are automatically generated without subsequent checking. Our pipeline provides a conservative means to do so. As our pipeline removes inconsistencies from NHCs data and estimates area of occupancy in an area slightly larger than the extent of occurrence of a species, compatible with IUCN red list assessments^{35,41}.

Methods

Species. We focused our analysis on 227 hyperdominant Amazonian tree species. The hyperdominant species are the most common tree species in Amazon, and together make up half of all trees with a diameter (dbh) over 10 cm²⁴. We chose only hyperdominant species to reduce the emergence of too many 'false absences' when plot data are interpolated into abundance maps. They present the largest probability of occurrence in the plots where they are present in the surrounding area.

Collections. Species collections were downloaded from GBIF (August 2017, www.gbif.org). We used data from the species' complete extent of occurrence to prevent deficiencies that are associated with SDMs based on a species' partial geographic range, such as under-prediction⁴². All individuals were assigned to species level; intraspecific levels were ignored.

Taxonomic names were checked with the Taxonomic Name Resolution Service (TNRS, <http://tnrs.iplant-collaborative.org/>). Although misidentification may represent a major problem in tree plots, we assume it is less severe in common species such as the hyperdominants; which are better represented in herbaria and more likely to be collected fertile²⁵. We assume that misidentification is within acceptable limits.

Collections cleaning pipeline. The cleaning pipeline consisted of a two-step process to remove inconsistencies from GBIF downloaded data (GBIF records). The first step consisted of removing all records with missing latitude, longitude or locality information (imprecise georeferences)³⁹ and all duplicates at 0.5-degree spatial resolution⁴⁰. With the GeoClean function from speciesgeocodeR R Package³⁸ we also removed coordinates assigned to capital cities, coordinates with latitude equal to longitude, coordinates equal to exactly zero; coordinates based on centroids of provinces, and corrected country references (cleaned GBIF records).

In the second step we used a kernel-density estimate function to remove spatial outliers from the cleaned GBIF data, assuming that these are misidentifications or incorrect coordinates not filtered by the step described above. This function calculates a fixed-bandwidth kernel-density estimate of the point process density function that produced the point patterns⁴³, using the density.ppp function from spatstat R Package⁴⁴ to generate a kernel-density estimate. Outliers were identified and removed based on the kernel-density values for each species coordinate, using a threshold based on a quantile function from stats R Package⁴⁵ (kernel-density estimate GBIF records).

The quantile threshold was set according to the number of Amazonian regions in which a species occurred, six in total as defined by ter Steege *et al.*²⁴. The quantile threshold was larger for species with narrow distribution (occurring in one to three Amazonian regions) and smaller for species with wide distribution (occurring in more than three Amazonian regions). As some hyperdominants are very widely distributed in Amazonia a larger quantile threshold cuts off too many occurrences, removing not only outliers, but also potential correct occurrence or entire occurrence clusters. Both steps reduced the number of species collection records (Appendix S4), and the predicted area of occupancy (Appendix S5).

Plot abundance data. Abundance maps were constructed using 1675 1-ha tree inventory plots well distributed across Amazonia (defined as the tropical rain forest of the Amazon basin and the Guyana Shield) from the Amazon Tree Diversity Network (ATDN) (<http://atdn.myspecies.info/>). All individuals with ≥ 10 cm diameter at breast height (dbh) were recorded within the plots²⁴. Because a relatively small number of collections from these plots have been deposited in herbaria, they constitute a dataset nearly independent from the NHCs.

Constructing abundance maps. Inverse distance weighting (IDW) interpolation was used to create abundance maps from the plot abundance data. First, Amazonia was divided into 2193 0.5-degree grid cells. We then constructed the inverse distance weighting (IDW) models based on relative abundance following ter Steege *et al.*²⁸. Then, the relative abundance (RA) for each cell was defined as $RA_i = n_i/N$, where: n_i = the number of individuals of species i , and N = the total number of trees. IDW models were based on the nearest 150 plots within a limit of 300 km distance. Each plot weight was calculated by taking the square root of the distance in degrees. The 150 plots that were taken into account ensured that within an area consisting of absence plots only, the species is predicted to be absent. In addition, the 3-degree distance limit causes the model to predict the absence of a species when no occurrence plots are present within a radius of 3 degrees. This setting is based on the notion that within a non-environmental model a species' extent of occurrence is restricted by dispersal limitation only⁴⁶. The maximum dispersal distance has been optimized to a 3-degree distance by determining the best match between the IDW maps and the Fisher's Alpha diversity map of all species⁴⁷.

Constructing presence-only SDMs using MaxEnt. We used MaxEnt version 3.3.3 k^{1,48}, to construct presence-only SDMs for all the 227 species. Data of 19 environmental variables were downloaded from WorldClim⁶. These included variables related to temperature and precipitation. Since collinearity, the non-independence of predictor variables, potentially leads to the wrong identification of relevant predictors for the model, we used the common Spearman's rank correlation coefficient threshold of $|\rho| > 0.7$ to identify correlated variables⁴⁹.

Subsequently, we selected least correlated variables ($|\rho| < 0.7$) based on biological relevance and their loadings in a principal component analysis (PCA). The PCA consisted of all environmental variables for all collection localities of the 227 species. For temperature, we selected isothermality, temperature seasonality, and maximum temperature of warmest month. For precipitation we chose annual precipitation, wettest month precipitation and driest month precipitation. All the environmental variables were cropped to the extent of the Neotropics⁴², and aggregated to a 0.5-degree spatial resolution, using the function 'mean' from R package 'raster'⁵⁰. We used precipitation and temperature variables to assess MaxEnt's predicted environmental suitability based on climate only. In the MaxEnt feature settings we excluded the product, threshold and hinge features given their lack of biological justification with the variables used^{34,36}.

Correcting for geographical sampling bias has been found to improve the predictive performance of MaxEnt¹⁴. Also, environmental bias can be assessed by environmental filtering, which improves MaxEnt discriminatory ability⁵¹. We produced a bias file to employ the target-group background method recommended by Phillips and Dudik⁵², an option which is implemented in MaxEnt. The bias file consisted of a binary raster grid based on all Amazon tree species collections²⁵, at each grid cell downloaded from GBIF, which reflects local survey effort. This is an essential step in the analysis, given MaxEnt's assumption that the occurrences are independently drawn from the unknown probability distribution of the species. Without a bias file, sampling bias could severely reduce models accuracy. We used the bias file to produce a background file according the efforts of collection. Finally we used a convex hull around cleaned occurrences (kernel-density estimate GBIF records) of each species to estimate their extent of occurrence *sensu*³⁵, plus a buffer of 300 km, equal to the buffer set for the IDW analysis, to crop the area of predicted environmental suitability^{29,41}. The latter is our predicted area of occupancy.

Data analysis. We compared collection presences and absences to IDW relative abundance to answer our first question whether NHCs are independent drawn from the unknown probability distribution. A binomial generalized linear model (logit regression) was used to determine if a significant positive relationship existed between the probability of being collected and predicted local relative abundance.

To answer the second question, how MaxEnt's predicted environmental suitability compares to IDW relative abundance, we first tested which species' MaxEnt maps were significantly different from random expectation with a bias corrected null-model⁵³. For each species, 99 null-models were generated by randomly drawing n collection localities without replacement from the same spatial grid as the environmental layers, with n being the number of geographically unique collections for that species. Using an upper one-sided 95% confidence interval, we determined the probability value of the observed AUC as calculated by MaxEnt against those generated by the null distribution. If the species' observed AUC value ranks 95 or above, the chance that a random set of n points could generate an equally good model is less than 5%, hence considered significantly different from random expectation. All species for which the SDM prediction did not deviate significantly from random expectation were excluded from further analysis.

Second, a Spearman Rank Correlation test was used to test the relationship between MaxEnt logistic output and IDW relative abundance at plot localities. Additionally, following VanDerWal *et al.*²⁹, we determined the linear 90th percentile quantile regression between the IDW relative abundance and MaxEnt logistic outputs at plot localities. The confidence intervals of the linear quantile regressions were calculated with the Markov chain marginal bootstrap method as suggested by Kocherginsky *et al.*⁵⁴. We computed the correlations and regressions for all plots separately, even if multiple plots were present in one grid square.

Third, we tested the predictive performance of MaxEnt and IDW. For MaxEnt, its logistic output was transformed into binary maps with a 10% training presence threshold. Although the maximum sum of sensitivity and

specificity is considered to be the best threshold method for presence-only SDMs by Liu *et al.*⁵⁵, we followed the advice of Merow *et al.*³⁴ to avoid measures with specificity because they are based on absences that are unknown in this analysis. Then we tested its sensitivity by calculating true positive rate of the binary maps against plot presence. That is, the fraction of the grid cells with a plot for which MaxEnt predicted the species correctly to be present. Finally we calculated the median predicted area of occupancy.

For IDW, its output was transformed into binary maps by converting the grids cells with RA >0 into 1. Last, naturally non-forested areas were excluded from the maps based on Soares-Filho *et al.*⁵⁶. We then calculated its output true positive rate against collections presences and absences. That is, the fraction of the grid cells with a collection for which the IDW relative abundance predicted the species correctly to be present. Finally we also calculated the median predicted area of occupancy for IDW.

All calculations and analyses were performed with R version 3.0.3³, including the R packages raster⁵⁰, rgdal⁵⁷, gstat⁵⁸, dismo⁵⁹, vegan⁶⁰, quantreg⁶¹, sp⁶², rJava⁶³ and SDMTools⁶⁴.

References

- Phillips, S. J., Anderson, R. P. & Schapire, R. E. Maximum entropy modeling of species geographic distributions. *Ecol. Modell.* **190**, 231–259 (2006).
- Elith, J. & Leathwick, J. R. Species Distribution Models: Ecological Explanation and Prediction Across Space and Time. *Annu. Rev. Ecol. Evol. Syst.* **40**, 677–697 (2009).
- Miller, J. Species Distribution Modeling. *Geogr. Compass* **4**, 490–509 (2010).
- Pearson, R. G. Species' distribution modeling for conservation educators and practitioners. *Lessons Conserv.* **3**, 54–89 (2010).
- Araújo, M. B. & Peterson, A. T. Uses and misuses of bioclimatic envelope modeling. *Ecology* **93**, 1527–1539 (2012).
- Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G. & Jarvis, A. Very high resolution interpolated climate surfaces for global land areas. *Int. J. Climatol.* **25**, 1965–1978 (2005).
- Kriticos, D. J. *et al.* CliMond: global high-resolution historical and future scenario climate surfaces for bioclimatic modelling. *Methods Ecol. Evol.* **3**, 53–64 (2012).
- Renner, I. W. & Warton, D. I. Equivalence of MAXENT and Poisson Point Process Models for Species Distribution Modeling in Ecology. *Biometrics* **69**, 274–281 (2013).
- Tobler, M., Honorio, E., Janovec, J. & Reynel, C. Implications of collection patterns of botanical specimens on their usefulness for conservation planning: an example of two neotropical plant families (Moraceae and Myristicaceae) in Peru. *Biodivers. Conserv.* **16**, 659–677 (2007).
- Haripersaud, P. P. P. Collecting biodiversity. (Utrecht University, 2009).
- ter Steege, H., Haripersaud, P. P., Bánki, O. S. & Schieving, F. A model of botanical collectors' behavior in the field: Never the same species twice. *Am. J. Bot.* **98**, 31–37 (2011).
- Beck, J., Böller, M., Erhardt, A. & Schwanghart, W. Spatial bias in the GBIF database and its effect on modeling species' geographic distributions. *Ecol. Inform.* **19**, 10–15 (2014).
- Phillips, S. J. *et al.* Sample selection bias and presence-only distribution models: implications for background and pseudo-absence data. *Ecol. Appl.* **19**, 181–197 (2009).
- Syfert, M. M., Smith, M. J., Coomes, D. A., Meagher, T. R. & Roberts, D. L. The Effects of Sampling Bias and Model Complexity on the Predictive Performance of MaxEnt Species Distribution Models. *PLoS One* **8**, e55158 (2013).
- Fourcade, Y., Engler, J. O., Rödder, D., Secondi, J. & Brooks, T. Mapping Species Distributions with MAXENT Using a Geographically Biased Sample of Presence Data: A Performance Assessment of Methods for Correcting Sampling Bias. *PLoS One* **9**, e97122 (2014).
- Kadmon, R., Farber, O. & Danin, A. Effect of Roadside Bias on the Accuracy of Predictive Maps Produced By Bioclimatic Models. *Ecol. Appl.* **14**, 401–413 (2004).
- Loiselle, B. A. *et al.* Predicting species distributions from herbarium collections: does climate bias in collection sampling influence model outcomes? *J. Biogeogr.* **35**, 105–116 (2008).
- Elith, J. *et al.* Novel methods improve prediction of species' distributions from occurrence data. *Ecography (Cop.)*. **29**, 129–151 (2006).
- Wisn, M. S. *et al.* Effects of sample size on the performance of species distribution models. *Divers. Distrib.* **14**, 763–773 (2008).
- Giovanelli, J. G. R. R., de Siqueira, M. F., Haddad, C. F. B. B. & Alexandrino, J. Modeling a spatially restricted distribution in the Neotropics: How the size of calibration area affects the performance of five presence-only methods. *Ecol. Modell.* **221**, 215–224 (2010).
- Merckx, B., Steyaert, M., Vanreusel, A., Vincx, M. & Vanaverbeke, J. Null models reveal preferential sampling, spatial autocorrelation and overfitting in habitat suitability modelling. *Ecol. Modell.* **222**, 588–597 (2011).
- Aguirre-Gutiérrez, J. *et al.* Fit-for-Purpose: Species Distribution Model Performance Depends on Evaluation Criteria – Dutch Hoverflies as a Case Study. *PLoS One* **8**, e63708 (2013).
- Fitzpatrick, M. C., Gotelli, N. J. & Ellison, A. M. MaxEnt versus MaxLike: empirical comparisons with ant species distributions. *Ecosphere* **4**, art55 (2013).
- ter Steege, H. *et al.* Hyperdominance in the Amazonian Tree Flora. *Science (80-)*. **342**, (2013).
- ter Steege, H. *et al.* The discovery of the Amazonian tree flora with an updated checklist of all known tree taxa. *Sci. Rep.* **6**, 1–15 (2016).
- Cámara-Leret, R., Tuomisto, H., Ruokolainen, K., Balslev, H. & Kristiansen, S. M. Modelling responses of western Amazonian palms to soil nutrients. *J. Ecol.* **1–15**, <https://doi.org/10.1111/1365-2745.12708> (2016).
- Graham, C. H. *et al.* The influence of spatial errors in species occurrence data used in distribution models. *J. Appl. Ecol.* **45**, 239–247 (2008).
- Steege, H. *et al.* Estimating the global conservation status of more than 15,000 Amazonian tree species. **9–11** (2015).
- VanDerWal, J., Shoo, L. P., Johnson, C. N. N. & Williams, S. E. Abundance and the Environmental Niche: Environmental Suitability Estimated from Niche Models Predicts the Upper Limit of Local Abundance. *Am. Nat.* **174**, 282–291 (2009).
- Calabrese, J. M., Certain, G., Kraan, C. & Dormann, C. F. Stacking species distribution models and adjusting bias by linking them to macroecological models. *Glob. Ecol. Biogeogr.* **23**, 99–112 (2014).
- Guillera-Arroita, G. *et al.* Is my species distribution model fit for purpose? Matching data and models to applications. *Glob. Ecol. Biogeogr.* **24**, 276–292 (2015).
- Lahoz-Monfort, J. J., Guillera-Arroita, G. & Wintle, B. A. Imperfect detection impacts the performance of species distribution models. *Glob. Ecol. Biogeogr.* **23**, 504–515 (2014).
- Lawson, C. R., Hodgson, J. A., Wilson, R. J. & Richards, S. A. Prevalence, thresholds and the performance of presence-absence models. *Methods Ecol. Evol.* **5**, 54–64 (2014).
- Merow, C., Smith, M. J. & Silander, J. A. A practical guide to MaxEnt for modeling species' distributions: What it does, and why inputs and settings matter. *Ecography (Cop.)*. **36**, 1058–1069 (2013).
- IUCN. IUCN Red List Categories and Criteria: Version 3.1. Gland, Switz. Cambridge, UK IUCN iv 32pp (2012).

36. Boucher-Lalonde, V., Morin, A. & Currie, D. J. How are tree species distributed in climatic space? A simple and general pattern. *Glob. Ecol. Biogeogr.* **21**, 1157–1166 (2012).
37. Maher, S. P., Randin, C. F., Guisan, A. & Drake, J. M. Pattern-recognition ecological niche models fit to presence-only and presence-absence data. *Methods Ecol. Evol.* **5**, 761–770 (2014).
38. Zizka, A. & Antonelli, A. speciesgeocodeR: An R package for linking species occurrences, user-defined regions and phylogenetic trees for biogeography, ecology and evolution. *bioRxiv* 32755 (2015).
39. Maldonado, C. *et al.* Estimating species diversity and distribution in the era of Big Data: To what extent can we trust public databases? *Glob. Ecol. Biogeogr.* **24**, 973–984 (2015).
40. Boyle, B. *et al.* The taxonomic name resolution service: an online tool for automated standardization of plant names. *BMC Bioinformatics* **13**, 14–16 (2013).
41. Syfert, M. M. *et al.* Using species distribution models to inform IUCN Red List assessments. *Biol. Conserv.* **177**, 174–184 (2014).
42. Raes, N. Partial versus full species distribution models. *Nat. a Conserv.* **10**, 127–138 (2012).
43. Diggle, P. J. A Kernel Method for Smoothing Point Process Data. *J. R. Stat. Soc. Ser. C (Applied Stat.)* **34**, 138–147 (1985).
44. Baddeley, A., Rubak, E. & Turner, R. Spatial point patterns: methodology and applications with R. (CRC Press, 2015).
45. Team, R. C. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2016. (2016).
46. Gaston, K. J. Geographic range limits: achieving synthesis. *Proc. R. Soc. B* **276**, 1395–1406 (2009).
47. ter Steege, H. *et al.* A spatial model of tree α -diversity and tree density for the Amazon. *Biodivers. Conserv.* **12**, 2255–2277 (2003).
48. Phillips, S. J., Dudík, M. & Schapire, R. E. A maximum entropy approach to species distribution modeling. *Proc. Twenty-First Int. Conf. Mach. Learn.* **83** (2004).
49. Dormann, C. F. *et al.* Collinearity: a review of methods to deal with it and a simulation study evaluating their performance. *Ecography (Cop.)* **36**, 27–46 (2013).
50. Hijmans, R. J. & van Etten, J. raster: Geographic Data Analysis and Modeling. *R package version 2*, 5–8 (2016).
51. Varela, S., Anderson, R. P., García-Valdés, R. & Fernández-González, F. Environmental filters reduce the effects of sampling bias and improve predictions of ecological niche models. *Ecography (Cop.)* **37**, 1084–1091 (2014).
52. Phillips, S. J. & Dudík, M. Modeling of species distributions with Maxent: New extensions and a comprehensive evaluation. *Ecography (Cop.)* **31**, 161–175 (2008).
53. Raes, N. & Ter Steege, H. A null-model for significance testing of presence-only species distribution models. *Ecography (Cop.)* **30**, 727–736 (2007).
54. Kocherginsky, M., He, X. M. & Mu, Y. M. Practical confidence intervals for regression quantiles. *J. Comput. Graph. Stat.* **14**, 41–55 (2005).
55. Liu, C., White, M. & Newell, G. Selecting thresholds for the prediction of species occurrence with presence-only data. *J. Biogeogr.* **40**, 778–789 (2013).
56. Soares-Filho, B. S. *et al.* LBA-ECO LC-14 Modeled Deforestation Scenarios, Amazon Basin: 2002–2050 (Oak Ridge National Laboratory Distributed Active Archive Center, Oak Ridge, TN, 2013). (2013).
57. Bivand, R., Keitt, T. & Rowlingson, B. rgdal: Bindings for the Geospatial Data Abstraction Library. R package version 0.8–16, <https://doi.org/10.1353/lib.0.0050> (2014).
58. Pebesma, E. & Graeler, B. gstat: spatial and spatio-temporal geostatistical modelling, prediction and simulation. R package version 1.0–19 (2014).
59. Hijmans, R. J., Phillips, S., Leathwick, J. & Elith, J. Species Distribution Modeling. Package ‘dismo’. *dismo: Species Distribution Modeling*. R package version 0.9–3, <http://CRAN.R-project.org/package=dismo> (2017).
60. Oksanen, A. J. *et al.* Package ‘vegan’. (2015).
61. Koenker, R. quantreg: Quantile Regression. R package version 5.05 (2013).
62. Pebesma, E. J. & Bivand, R. sp: classes and methods for spatial data in R. R package version 1.0–15 (2014).
63. Urbaneck, M. S. rJava: low-level R to Java interface. R package version 0.9–6 (2013).
64. VanDerWal, J., Falconi, L., Januchowski, S., Shoo, L. & Storlie, C. SDMTTools: Species Distribution Modelling Tools: Tools for processing data associated with species distribution modelling exercises. R package version 1.1–20 1 (2014).

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Author Contributions

H.t.S. conceived the study. H.t.S., N.R., S.D.I.J. and V.H.F.G. designed the study. V.H.F.G. carried out the GBIF data collection. V.H.F.G. and S.D.I.J. carried out the analyses and wrote the R scripts. All co-authors contributed to checking and re-checking of the species list and all co-authors contributed to the writing.

Additional Information

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