

Accepted refereed manuscript of:

Gonzalez Diaz P, Jump A, Perry A, Wachowiak W, Lapshina E & Cavers S (2017) Ecology and management history drive spatial genetic structure in Scots pine, *Forest Ecology and Management*, 400, pp. 68-76.

DOI: [10.1016/j.foreco.2017.05.035](https://doi.org/10.1016/j.foreco.2017.05.035)

© 2017, Elsevier. Licensed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International
<http://creativecommons.org/licenses/by-nc-nd/4.0/>

Title

Ecology and management history drive spatial genetic structure in Scots pine

Authors and affiliations

Patricia González-Díaz^{a,b}, Alistair S. Jump^{a,c}, Annika Perry^b, Witold Wachowiak^{b,d}, Elena Lapshina^e and Stephen Cavers^b

^a Biological and Environmental Sciences, Faculty of Natural Sciences, University of Stirling, Stirling, FK9 4LA, UK.

^b Centre for Ecology and Hydrology Edinburgh, Bush Estate, Penicuik, Midlothian EH26 0QB, UK.

^c CREAF (Centre de Recerca Ecológica i Aplicacions Forestals), Campus UAB, Edifici C. E-08193, Belaterra (Barcelona), Spain.

^d Institute of Dendrology, Polish Academy of Sciences, Parkowa 5, 62-035 Kórnik, Poland.

^e Yugra State University, Centre for Environmental Dynamics and Climate Change, Khanty-Mansiysk, 628012, Russia.

Corresponding author

Patricia González-Díaz: patricia.gonzalezdiaz@stir.ac.uk

Keywords

Pinus sylvestris, spatial genetic structure, genetic diversity, forest management, life stages.

Accepted for publication in *Forest Ecology and Management* by Elsevier.

Abstract

Forest management practices that remove trees from stands can promote substantial changes in the distribution of genetic diversity within and among populations at multiple spatial scales. In small and isolated populations, elevated inbreeding levels might reduce fitness of subsequent generations and threaten forest resilience in the long term. Comparing fine-scale spatial genetic structure (SGS) between life stages (e.g. adult and juvenile cohorts) can identify when populations have undergone disturbance, even in species with long generation times. Here, we studied the effects of historical and contemporary forest management, characterized by intense felling and natural regeneration respectively, on genetic diversity and fine-scale SGS in adult and juvenile cohorts. We examined fragmented Scots pine (*Pinus sylvestris* L.) stands in the Scottish Highlands, and compared them with a remote, unmanaged stand. A total of 777 trees were genotyped using 12 nuclear microsatellite markers. No difference was identified in allelic richness or gene diversity among stands or life stages, suggesting that historical and contemporary management have not impacted levels of genetic variation. However, management appears to have changed the spatial distribution of genetic variation. Adult genotypes from managed stands were more spatially structured than in the unmanaged stand, a difference mediated by contrasts in tree density, degree of fragmentation of stands at the time of establishment and rate of gap creation. Surprisingly, juveniles were less spatially structured than adults in the managed stands, suggesting an historical erosion of the structure of the adult cohort but contemporary recovery to natural dynamics, and indicating a high capacity of the species to recover after disturbance. Here we showed that using the spatial component of genetic diversity can help to detect both historical and contemporary effects of disturbance in tree populations. Evaluation of successional change is important to adequately detect early responses of tree populations to forest management practices. Overall, our study suggests that combining sustainable management with forest conservation practices that ensure larger effective population sizes is key to successfully maintaining genetic diversity in Scots pine.

1. Introduction

A prolonged history of forest exploitation based on the harvesting of trees has resulted in widespread modification of Europe's forests, impacting genetic diversity within and among populations (FAO, 2014). Currently, over 70% of European forests (representing some 15% of European forest area) are subject to a management plan or its equivalent (Forest Europe, 2015) but, despite a substantial shift toward sustainable practices over the past 25 years (FAO, 2015), the consequences of historical management practices such as extensive felling on the distribution of genetic diversity in tree species remain largely uncertain. Genetic diversity plays an essential role in underpinning forest resilience by facilitating evolutionary processes, and it is key in forest responses to disturbances, such as habitat loss, fragmentation or pathogen attack (Schaberg et al., 2008; Cavers and Cottrell, 2014). Consequently, understanding how historical and contemporary forest management have shaped patterns of genetic diversity allows evaluation of the potential resilience of European forests and informs the development of adaptive management plans.

The impact that tree removal can have on population genetics has been addressed through exploration of levels of neutral genetic variation, revealing changes in gene frequencies (Schaberg et al., 2008) and loss of alleles (Adams et al., 1998; Rajora et al., 2000; Kettle et al., 2007; Ortego et al., 2010), yet many studies have failed to detect significant effects (Bradshaw, 2004; García-Gil et al., 2015; Rajora and Pluhar, 2003; Schaberg et al., 2008; Young et al., 1996). Some authors attribute the lack of effect to the long generation time in trees, because changes in genetic diversity after disturbance may take many generations (Lowe et al., 2005). However, changes in tree distribution and age structures can alter the spatial organisation of genetic variation, even when overall levels of variation are maintained, allowing us to explore the genetic legacy of forest management (Piotti et al., 2013; Sjölund and Jump, 2015).

In naturally regenerated tree populations, genotypes are not distributed randomly. Typically, individuals become less genetically similar as the distance between them increases (Jump and Peñuelas, 2007; Paffetti et al., 2012; Vekemans and Hardy, 2004), causing a phenomenon known

as spatial genetic structure (SGS). Restricted dispersal results in offspring being more likely to establish close to the mother tree (Jump et al., 2012; Pandey et al., 2012). Consequently, the pollen and seed dispersal mechanism will strongly influence the extent and magnitude of SGS within a species. For example, plants with animal dispersed pollen usually show greater SGS than those with wind dispersed pollen (Vekemans and Hardy 2004). Furthermore, individual density is usually inversely correlated with SGS. For example, the extent of SGS in low density populations of *Acer pseudoplatanus* is nine times greater than in high density populations (Vekemans and Hardy 2004).

The ecological determinants of SGS (such as recruitment frequency, seed and pollen dispersal distance, and individual density) are commonly modified by forest management practices that remove trees. Consequent changes in SGS may alter local mating patterns and the distribution of genetic diversity in subsequent generations (Smouse and Peakall, 1999). Furthermore, different forest management practices, such as felling, coppicing or thinning, will differentially impact selection of individuals and seedling establishment potentially leading to a broad range of genetic impacts (Cottrell et al., 2003; Paffetti et al., 2012; Piotti et al., 2013; Sjölund and Jump, 2015). Distinguishing the effects of forest management on SGS is, therefore, a challenging task.

SGS of plant populations is dynamic and can change across life stages. In individuals that reproduce sexually, seedlings might be affected by compensatory mortality and competitive thinning, post dispersal, thereby altering spatial distribution patterns with age (Ng et al., 2004). Most studies have found greater SGS in early regeneration stages than in mature individuals (González-Martínez et al., 2002; Hardesty et al., 2005; Ng et al., 2004; Soto et al., 2007; Troupin et al., 2006). The successional component of SGS (e.g. comparing SGS between adult and juvenile cohorts) has mainly been studied in order to understand the natural development of SGS (Berens et al., 2014; González-Martínez et al., 2002; Jones and Hubbell, 2006). Such changes in SGS have rarely been used to assess the influence of forest management practices (but see Jones et al., 2006; Leclerc et al., 2015; Troupin et al., 2006).

This study focuses on the remaining fragmented Scots pine (*Pinus sylvestris* L.) forests of the Scottish Highlands (known as Caledonian pine forests), which are believed to be the only native pine forests in the UK. These fragmented remnants represent a valuable system in which to study the impacts of historical forest management practices because numerous records of management history exist. To understand the effects of historical and contemporary forest management practices, we investigated genetic diversity and fine-scale SGS in adult and juvenile cohorts in two native managed pine forests and compared these with a remote, unmanaged stand. We selected two life stages that were established in distinct periods with contrasting forest management systems: (1) adult trees that established during 19th Century, characterised by high browsing pressure by deer and after a period of intense felling (hereafter historical management); and (2) juveniles that established during the last two decades, characterised by conservation policies promoting natural regeneration (hereafter contemporary management). Specifically we sought to determine: 1) did historical management practice impact genetic diversity and SGS – comparing mature managed and unmanaged stands? and 2) how has contemporary management practice affected diversity and SGS – comparing adults and juveniles from managed stands? We hypothesised that in the absence of effects of historical management, mature managed stands would display similar values of genetic diversity and SGS as those in an unmanaged stand, while in the absence of effects of contemporary management, stronger SGS would be found in the juvenile stages, and similar values of genetic diversity will be evident in both juvenile and adult cohorts.

2. Material and methods

2.1. Study species

Scots pine is a wind-pollinated outcrossing conifer and is the most widely distributed pine species in the world, with a range that spans Eurasia, from the Arctic circle in Norway in the north to the south of Spain and south of Turkey and from the west coast of Scotland to the far east of Russia (Carlisle and Brown, 1968). Populations from southern Europe, Scotland and Asia Minor

generally represent isolated occurrences. In Scotland this species occurs at the western limit of its global distribution and constitutes the iconic species of the Caledonian pine forest. Scots pine is typically a pioneer species (together with birch and aspen) that readily regenerates after natural or human disturbances, if competition and grazing pressure are low (Mátyás et al., 2004). It grows well on most soils, nevertheless, due to shade and competition intolerance, it is often restricted to poor soils (Steven and Carlisle, 1959). It is a monoecious species, and female flowering can start at the age of 15 to 30 years, in open to closed stands respectively (Mátyás et al., 2004). Pollen movement is predominantly over tens of metres within a stand (Robledo-Arnuncio et al., 2004b), but it may reach 100 km (Robledo-Arnuncio, 2011). Seeds are primarily wind and gravity dispersed, and typically travel up to 100 metres (McVean, 1963).

2.2. Study sites and history of forest management

From a peak distribution around 6,000 years ago, Scots pine in Scotland has been in decline for millennia, with a major retreat 4,000 years ago, initially attributed to a climate shift to wetter conditions (Bennett, 1984), although human and grazing pressures may have also played a significant role (Tipping et al., 2008). The exploitation and reduction in Scots pine extent has been particularly intense from the 18th Century onwards (Hobbs, 2009), mainly characterized by felling and selective logging to provide construction timber (Smout, 2003). The general decrease in forest extent, together with poor natural regeneration in the Caledonian pine forest (due to extensive browsing pressure by deer and sheep), kept this forest at low tree density for many years (McVean, 1963) and has strongly suppressed regeneration during the last 200 years (Steven and Carlisle, 1959). During the last few decades, however, forest management has moved to protect and expand the remaining Caledonian pine forest (Forestry Commission, 2016).

We selected two study sites in Scotland, Abernethy (57°20'N, 3°61'W) and Glen Affric (57°15'N, 5°00'W). Nowadays, these sites constitute some of the largest ancient pine forest in Scotland covering areas of 2452 ha and 1532 ha, respectively (Mason et al., 2004). In each site, an old open native stand was selected, where trees are expected to have been established through

169 natural regeneration of native provenance. Hereafter these stands will be referred to as managed
170 stands. The fire regime in the UK is largely human driven (Davies et al., 2008), but tree mortality
171 through large fires is uncommon in Scotland. In addition, wind-blow and snow can cause some
172 casualties through the year, and fungi and insects will be minor effects. However, intense forest
173 disturbance in recent centuries can be attributed mainly to forest management practices.

174
175 The study site in Abernethy is located at 370 m a.s.l., with south westerly prevailing winds and
176 densities of 160 stems ha⁻¹. Stand composition is formed by Scots pine, with presence of *Juniperus*
177 *communis*. The understory is predominantly *Calluna vulgaris*, *Vaccinium myrtillus* and small
178 patches of *V. vitis-idaea*. Historical exploitation at Abernethy has taken place over millennia and
179 high felling and browsing pressure during the 18th Century are reflected in the progressive
180 contraction of the extent of Abernethy forest in historical maps from 1750 until 1830 (Smout et
181 al., 2005, Summers et al. 2008). By 1858, the forest was represented only by scattered trees in the
182 study site and followed by enclosure of the forest as deer forest occurred in 1869 (O’Sullivan,
183 1973). In the 1980s the area was designated a National Natural Reserve. Seasonal grazing by
184 sheep was stopped in 1990 and deer fences were removed (Beaumont et al., 1995). Since then,
185 culling of deer has kept the population stable and compatible with forest regeneration. By 1992
186 the percentage of seedlings with evidence of browsing had fallen from 72% to 43% with an
187 increase of 20% in the number of established seedlings and saplings (Beaumont et al., 1995).

188
189 The study site in Glen Affric is located at 260 m a.s.l., south west of Loch Affric, where the
190 prevailing winds are south westerly, and stand density is 103 stems ha⁻¹. Stand composition is
191 dominated by Scots pine and the vegetation layer is predominantly *C. vulgaris* with small patches
192 of *V. vitis-idaea* and *V. myrtillus*. Evidence from pollen records from west Glen Affric, where our
193 stand is located, show a sustained low tree cover around these sites for several thousand years as
194 a result of prolonged human impact, with the recent expansion of the forest when the present tree
195 cohort developed around 1880 (Shaw, 2006). Historical documents report felling of trees during
196 the 18th and 19th Centuries (Smout et al., 2005) with the decline evident in pollen records.

Following a period of intensive sheep and deer grazing in the late 20th Century a major effort was made to protect and restore the remaining native pine forest (Bain, 2013). Glen Affric was initially declared as a Caledonian Forest Reserve in 1961 by the Forestry Commission (Bain, 2013) and later, in 1984, a National Natural Reserve.

To compare our heavily managed stands with an unmanaged case, and since unmanaged stands do not exist in Scotland, pre-existing samples from a boreal site in Western Siberia were used (60°54'N, 68°42'E). These samples were taken from within a continuous population with extensive areas of natural forest, with a stand density of 470 stems ha⁻¹. These forests have never been altered by humans, but are subject to regular natural disturbance by fire on roughly 50 year timescales. In these boreal forests, competition forces Scots pine to forest edges and onto poor quality sites such as sandy soils or bogs, and it is outcompeted on better soils by *Pinus sibirica*, *Larix sibirica* and *Populus tremula*. As a result even mature individuals may be small. Hereafter this stand will be referred to as the unmanaged stand.

In Scots pine, genetic variation is partitioned predominantly within rather than among populations, and levels of within-population genetic diversity across the range of Scots pine are similarly high (Wachowiak et al., 2014, 2011). Previous studies of diversity across the range of this species have shown that genetic differentiation among even distant populations of Scots pine is low (Naydenov et al., 2007; Provan et al., 1998; Prus-Glowacki and Stephan, 1994; Wang et al., 1991) but see (Forrest, 1982; Prus-Glowacki et al., 2012). Some authors attribute this homogeneity to common ancestry, as well as extensive gene flow (Chybicki et al., 2008) and lack of major physical barriers (Naydenov et al., 2007). As absolute genetic diversity levels in the managed and unmanaged stands are of similar magnitude, and the physical capacity for gene movement should be similar in each, we can assume that the primary driver of genetic structure will have been the presence or absence of significant human intervention. Therefore, this comparison can be informative regarding the processes that are likely responsible for the observed spatial pattern of genetic diversity at fine scales.

2.3. Sample collection, life stages and stand structure

We selected stands of 200 m × 200 m in Abernethy and Glen Affric, respectively. Sampling strategy was designed to account for short distance classes in order to detect fine-scale SGS, choosing individuals randomly and assuring sufficient numbers of pairwise comparisons in each distance class, as recommended by Cavers et al. (2005). We sampled needles from two life stages, juveniles and adults. Sample size per stand in each life stage varied from 131 to 181 (Table 1). All individuals were mapped using a GARMIN 62s handheld GPS and diameter was measured at breast height (d.b.h.). The d.b.h. was used as a proxy of age, defining juveniles as individuals with d.b.h. below 10 cm. Existing data from trunk cores from nearby adult pines in Abernethy (Summers et al., 2008) were used to calibrate the relationship between d.b.h. and age.

The unmanaged study site was sampled in three sub-stands of 50 x 50 m along a linear transect of 300 m, which were combined to give a single stand sample for subsequent analysis. All sampled individuals were mapped, measured for d.b.h. and tree height classified as short (<2m) or tall (>2m). Juveniles were defined as short individuals with d.b.h. below 10 cm. Sample size in each life stage varied from 57 to 73 (Table 1). Thirty random trunk sections from adult pines were taken from the unmanaged site to calibrate the d.b.h.-age relationship. We evaluated the relationship between d.b.h. and tree age, and whether this relationship varied among sites using a linear model in R 3.0.1 (R Core Team, 2013). We chose d.b.h. as the response variable and tree age and site (Abernethy and unmanaged) were the predictor variables. The interaction between the predictor variables was tested and compared with a model without interactions by using the Akaike Information Criterion.

2.4. Microsatellite analyses

Total genomic DNA was extracted from 50 mg silica gel dried needles using QIAGEN DNeasy 96 Plant Kit (QIAGEN Ltd. Crawley, UK) following the manufacturer's protocol. All individuals were genotyped at twelve nuclear microsatellite markers (SSR): psyl2, psyl16, psyl17, psyl36,

psyl42, psyl44, psyl57 (Sebastiani et al., 2011), SPAC7.14, SPAC12.5 (Soranzo et al., 1998), PtTX4001, PtTX4011 (Auckland et al., 2002) and SsrPt_ctg4698 (Chagné et al., 2004), combined in two multiplexes of six loci each. Multiplex 1 consisted of primers psyl2, psyl17, psyl42, psyl44, PtTX4001 and PtTX4011 at concentrations of 3 µl, 2 µl, 2 µl, 2 µl, 3 µl and 2 µl respectively. Multiplex 2 consisted of primers psyl16, psyl36, psyl57, SPAC7.14, SPAC12.5 and SsrPt_ctg4698 at concentrations of 2 µl each. Reactions were carried out in a final volume of 10 µl with 1X of QIAGEN Type-it Multiplex PCR Master Mix, 1 µM of each multiplex and 25 ng of template DNA. Annealing temperature for both multiplexes was 56°C. Polymerase chain reactions (PCR) were performed in Veriti™ Thermal cycler (Applied Biosystems, Bleiswijk, Netherlands), with the following programme: 1 cycle at 95°C for 4 min followed by 35 cycles at 95°C for 45 s, 56°C for 45 s, 72°C for 45 s, and a final step at 72°C for 5 min. PCR products were analysed by DNA Sequencing and Services, Dundee, UK, using an Applied Biosystems 3730 DNA Sequencer with reference to a LIZ 500 size standard. Fragment analysis results were scored using GENEMARKER V.2.6.0. (SoftGenetics, State College, PA, USA). FLEXIBIN (Amos et al., 2007) was used to check discrete classes of raw allele sizes and MICRO-CHECKER (Van Oosterhout et al., 2004) to check genotyping errors and null allele frequencies. Several markers showed evidence of null alleles at very low frequencies (maximum frequency of 0.04, data not shown), which is far below to the threshold at which null alleles can result in a significant underestimate of expected heterozygosity, estimated as 0.2 (Belletti et al., 2012; Chapuis and Estoup, 2007). Therefore, all markers were kept for further analysis.

2.5. Genetic diversity and spatial genetic structure analysis

Genetic diversity estimators within stands and life stages were estimated using FSTAT 2.9.3.2 (Goudet, 1995): mean number of alleles per locus (A), rarefied allelic richness (A_R) (rarefied to 57 individuals for each stand and life stage), expected heterozygosity (H_E) and inbreeding coefficient (F_{IS}). We conducted ANOVAs to test for differences in A , A_R , and H_E between stands and life stages in R 3.0.1 (R Core Team 2013). We calculated F_{ST} among stands and life stages in ARLEQUIN v3.5 (Excoffier and Lischer, 2010), and the differentiation index D (Jost, 2008)

implemented in the R package DEMETics (Gerlach et al., 2010). In both cases, significance values were determined for a 5% nominal level after Bonferonni correction. F_{ST} estimates differences in allele frequencies among stands, whereas differentiation index D measures the fraction of allelic variation among them.

We implemented fine scale SGS analyses in SPAGeDi 1.4b (Hardy and Vekemans, 2002). In order to test for significance in genetic relatedness, the Kinship coefficient of Loiselle et al. (1995) (F_{ij}) was estimated as $F_{ij}=(Q_{ij}-Q_m)/(1-Q_m)$, where Q_{ij} is the probability of identity in state for random gene copies from two individuals i and j , and Q_m is the average probability of identity by state for gene copies coming from random individuals from the sample. A regression between the Kinship coefficient F_{ij} and the logarithm of pairwise geographic distances of individuals was computed (b_F). Standard errors of the regression slope were computed using a jackknife procedure over loci. The significance of the slope of the regression was tested using 10,000 permutations of locations among individuals. To visualize the SGS, we plotted average pairwise estimates of genetic relatedness as a function of distance to generate spatial autocorrelograms. Analyses were conducted for each stand and life stage separately across the full distance range. SGS_{MAX} was also calculated for each stand and life stage, which is the greatest distance at which the Kinship coefficient of a given distance class $F(d)$ is significant at $p<0.05$ (Jump et al., 2012). We also calculated the Sp statistic, as suggested by Vekemans and Hardy (2004), to allow comparability among stands and life stages with other studies. The Sp statistic was determined as $-b_F/(1 - F_1)$, where b_F is the regression slope of kinship coefficient estimate (F) on distance classes and F_1 is the kinship coefficient for adjacent individuals in the first distance interval.

Because the number of pairs within each distance class should ideally exceed 50 pairs of individuals, we set the distance intervals of at least 10 metres (Cavers et al., 2005; Jump and Peñuelas, 2007). Overall, we established 10 distance classes for the managed stands (0-10, 10-20, 20-30, 30-40, 40-50, 50-60, 60-70, 70-80, 80-90, 90-100), and 8 distances classes in the unmanaged stand (0-10, 10-20, 20-30, 30-60, 60-70, 70-80, 80-90, 90-100). Distance classes

between 30 and 60 metres were combined in the unmanaged stand to ensure sufficient numbers of pairs in the class. We also tested other distance class options and longer final distances up to 200 metres, and found they revealed similar and no more informative results. In addition, in the unmanaged stand, analysis of each sub-stand was also conducted separately, and the same results were obtained.

3. Results

3.1. Stand structure

Tree diameter distribution for managed stands was bimodal, with the highest frequencies for juvenile individuals at diameters between 0 and 10 cm (Fig. 1). A gap of adult individuals with diameter classes between 10 to 30 cm and 10 to 25 cm occurred in Abernethy and Glen Affric, respectively (Fig. 1). Contrastingly, tree diameter distribution in unmanaged stand was more skewed towards smaller diameters. There was no gap in the distribution in this case, instead there was a gradual decrease in the numbers of individuals with increasing diameter class (Fig. 1).

We found that d.b.h. was dependent on age and site ($F=29.85$, $R^2=0.31$), showing strong differences among age ($t=3.81$, $p<0.001$), and among sites ($t=-6.03$, $p<0.001$). However, we did not find significant interactions between age and study site (Fig. 2). The relationship between d.b.h. and age suggested that differences in age profiles in the two sites were smaller than differences in tree size (e.g. trees with different d.b.h. could have a similar age).

3.2. Genetic diversity

Genetic diversity parameters did not significantly differ between managed and unmanaged stands (Table 1). Among the twelve nuclear loci analysed, the number of alleles (A) in the managed stands ranged from 3 to 31 and 4 to 29 per locus for Abernethy and Glen Affric respectively for both life stages combined (multilocus average of 9.92 for each site). A ranged from 3 to 31 in the unmanaged stand, with a multilocus average of 9.83 again for both life stages combined. For rarefied allele richness (A_R) in the managed stands, multilocus estimates obtained mean values of

8.99 and 8.83 for Abernethy and Glen Affric respectively and 8.95 for the unmanaged stand both life stages combined, based on a minimum number of 126 individuals. Expected heterozygosity levels (H_E) showed multilocus estimates of 0.58 in Abernethy and 0.56 in Glen Affric, and similar values of 0.58 for the unmanaged stand for both life stages combined (See Table 1 for genetic diversity estimators on each site and life stage and Appendix A, Table A1, for detailed information of each microsatellite). Neither A , A_R or H_E significantly differed between managed vs. unmanaged stands (all p -values > 0.05). However, some differences appeared in the inbreeding coefficient (F_{IS}) which was significant and higher for both managed stands, indicating significant departure from Hardy–Weinberg equilibrium, whereas it was not significant for the unmanaged stand (Table 1). F_{ST} values indicated low but significant differences among the two managed stands ($F_{ST}=0.004$, $p<0.001$), and higher differences when comparing them with the unmanaged stand ($F_{ST}=0.03$ and $F_{ST}=0.04$, $p<0.001$, for Abernethy vs. unmanaged and Glen Affric vs. unmanaged respectively), indicating that despite remarkably similar overall levels of genetic diversity, their genetic composition differs to some extent.

When comparing life stages within stands, neither A , A_R or H_E significantly differed (all p -values > 0.05). F_{ST} values indicated no significant differences among life stages in Abernethy and the unmanaged stand, however low but significant F_{ST} occurred among life stages in Glen Affric. In agreement, differentiation index D showed the same pattern, although values were consistently larger (See Appendix A, Table A2).

3.3. Spatial genetic structure

We found significant SGS in all managed stands for both life stages which extended up to 40 metres further than the unmanaged stand (Table 1 and Fig. 3). The kinship coefficient for the first distance class $F_{(1)}$ and the Sp statistic also reflected the relationship between the extent of SGS and historical management, which was larger for managed than for unmanaged stands (Table 1).

When comparing SGS among life stages within stands, both SGS_{MAX} and $F_{(I)}$ were larger for adult than for juvenile stages in the managed stands (e.g. SGS_{MAX} extended up to 20 metres further in adults than juveniles) (Table 1 and Fig. 3). In contrast, SGS was larger for juveniles than for adults for the unmanaged stand, with significant SGS only at distances of less than 10 metres in the juvenile stage (Table 1 and Fig. 3). In the managed site of Glen Affric, we found that at 50-80 m trees were less genetically similar than expected compared with a random distribution of genotypes (see significant negative values of Kinship coefficient at distances between 50 and 80 metres in Glen Affric in Fig. 2). The minimum number of pairwise comparisons per distance class in the managed stands for each life stage was 106 individuals, whereas it was 63 individuals in the unmanaged stand. The Sp values did not reflect the same relationship between the extent of SGS with contemporary management as SGS_{MAX} and $F_{(I)}$ did. Thus, of the managed stands, Sp value was not significantly different between adults and juveniles in Abernethy, whereas it increased from adults to juveniles in Glen Affric (Table 1).

4. Discussion

We found two main results: 1) although overall levels of genetic diversity were strikingly similar, more extensive spatial structuring of genetic diversity was found in the mature managed stands when compared with the unmanaged one; 2) in contrast to expectations, a reduced extent of spatial genetic structure was found in the early stages of regeneration of the managed stands compared with the adult cohorts, again despite no difference in overall levels of genetic diversity between life stages. These patterns suggest that both historical and contemporary management can significantly alter spatial genetic structure of Scots pine. Here, we combine ecological information with historical data on the stands to better understand the mechanisms that are likely responsible for these differences in spatial genetic structure.

4.1. Impact of historical forest management practices

Notable differences in size profiles appeared between managed and unmanaged stands, (e.g. mean d.b.h. generally bigger in managed stands (Fig. 1)). However, the d.b.h.-age relationship was

different among managed and unmanaged stands (Fig. 2), linked to the growth-retarding effect of the bog habitat of the latter. Hence, the contrast in age profiles –a more important comparison for SGS analysis– was much smaller than for size profiles (e.g. small trees from the unmanaged stand often had similar ages to much larger trees from the managed one). The age profile of the stands was strongly reflective of their distinct histories, with large, old trees present in the managed sites plus a pulse of recent regeneration, whilst a much wider range of ages was present in the unmanaged one, with fewer very old trees. The structure in the unmanaged site is likely to reflect the natural fire disturbance dynamics to which it is exposed. These dynamics are likely in turn to affect genetic structure, with a higher turnover in the unmanaged stand –shown by the diverse, but generally young age profile– indicating a higher potential for gene dispersal and therefore erosion of spatial structure.

Genetic diversity of both mature managed sites, as indicated by allelic richness and expected heterozygosity, did not differ significantly from the unmanaged stand but instead was remarkably similar (e.g. H_E : 0.57-0.59 vs. H_E : 0.58, respectively). Although average diversity levels were lower than those reported in mainland European populations of Scots pine using nuclear SSR (H_E : 0.62-0.85) (Scalfi et al. 2009; Naydenov et al. 2011; Nowakowska et al. 2014; García-Gil et al. 2015) differences might be explained by the number of markers used and their specific levels of polymorphism. Thus, for example, selecting two of the three markers used by Scalfi et al. (2009), SPAC 7.41 and SPAC 12.5, the mean value of genetic diversity in our study (0.9) would be higher than previously reported. Also, the markers with the lowest values of diversity in our study, psy144 and psy12, had very similar low values in mainland European populations (Sebastiani et al., 2011) (see Appendix A, Table A1). Previous studies in Scottish populations of Scots pine have also reported relatively high levels of genetic variation using other molecular markers (Forrest, 1982, 1980; Kinloch et al., 1986; Provan et al., 1998; Sinclair et al., 1998; Wachowiak et al., 2013, 2011).

High levels of genetic variation at the population level suggests that effective population size has been sufficiently high to restrict effects of genetic drift despite intensive management and geographical isolation of populations. Scots pine is a wind-pollinated tree with wind-dispersed seed, and achieves high levels of gene flow by: (1) long seed wings, up to four times as long as the seed (Steven and Carlisle, 1959), (2) low seed mass (Castro, 1999) (here 2.9 to 12.64 mg), on average smaller than other pine species (9.1 to 233 mg) (Vander Wall, 2003), and (3) extensive pollen flow, from 17-22 m (Robledo-Arnuncio et al., 2004b) and up to 100 km in small fragments (Robledo-Arnuncio, 2011) (similar to other wind-pollinated tree species). Therefore, it appears that gene flow has been sufficient to prevent erosion of genetic diversity. F_{IS} values, an indirect measure of inbreeding, were not high in the managed sites although they were significantly higher than in the unmanaged site (0.05-0.06 vs. 0.01 respectively), suggesting that although gene flow has prevented loss of genetic diversity at the population level, fine scale alterations to gene flow might have taken place. Drastic reduction of population sizes can induce higher rates of selfing and mating between relatives (Robledo-Arnuncio et al., 2004a). The small size of the population at the time of establishment of the current adult cohorts, as indicated by historical data (Shaw, 2006; Summers et al., 2008), might explain this pattern.

Consistent differences in SGS were found in the mature managed stands which showed greater extent and magnitude of structure, as indicated by SGS_{MAX} up to 40 metres and higher $F_{(I)}$, compared with the unmanaged one. The extent of SGS of the mature managed stands was also larger than the values reported for Scots pine (Chybicki et al., 2008) and other *Pinus* species, which typically had values below 15 metres (De-Lucas et al., 2009; González-Martínez et al., 2002; Jones et al., 2006; Marquardt and Epperson, 2004; Parker et al., 2001; Troupin et al., 2006; Williams et al., 2007). It should be noted, however, that SGS estimates in Parker et al. 2001 and Jones et al. 2006 were based on allozyme markers, and the need for caution when comparing SGS extent with different molecular markers has been previously highlighted (Jump and Peñuelas, 2007).

Values of SGS extent more comparable to those in our managed stands were observed in fragmented populations of *Pinus pinaster* (~ 20 metres) (De-Lucas et al., 2009). The high values of SGS_{MAX} in the managed stands are likely a consequence of the drastic reductions in the number of seed and pollen donors, which are two of the main drivers of SGS (e.g. due to felling practices). The larger extent of SGS observed in Glen Affric may arise from local differences in historical management, with a prolonged limited tree cover due to human activities (Shaw, 2006), which is also reflected in the lower density of the site. The very short spatial scale of genetic structure in the mature unmanaged stand was remarkably similar to that in undisturbed continuous populations of *P. pinaster* which displayed either weak or no relatedness, with maximum values of SGS_{MAX} of 10 metres (De-Lucas et al. 2009). As these populations have contrasting local contexts, the unmanaged stand being part of the extensive core Eurasian population whereas the undisturbed population of *P. pinaster* being a distributional edge population, the similarity in SGS values observed seems likely to be due to their common unmanaged state. Therefore, it seems clear that tree density, degree of fragmentation of stands at the time of establishment and rate of gap creation play a major role in the formation of SGS in populations. Sp values for the mature managed stands (0.0045 and 0.0098) were remarkably higher than for the non-managed stand (-0.0006). Similarly, De-Lucas et al. (2009) found differences in the Sp values between fragmented and continuous populations of *P. pinaster*, although they were generally higher than the values reported in this study.

4.2. Impact of contemporary forest management practices

In the managed stands, there were no differences among life stages in the levels of allelic richness or gene diversity, suggesting contemporary management has not impacted genetic variation. However, we found higher relatedness – as higher SGS intensity and extent – in adults than in juveniles, with a greater discrepancy in the Glen Affric site. In contrast, the unmanaged site had stronger relatedness in the juvenile stage than in adults, as is usually found in natural tree populations. Natural populations often show greater SGS in the early stages of regeneration, due to the higher spatial aggregation of trees (Rozas et al., 2009; Szwagrzyk and Czerwczak, 1993).

This pattern has been reported in other species of *Pinus* (González-Martínez et al., 2002), in *Quercus* (Hampe et al., 2010), tropical trees (Hardesty et al., 2005; Ng et al., 2004) and other plant species (Yamagishi et al., 2007). Nevertheless, a few studies have found greater SGS in adult life stages, such as in *Jacaranda copaia* (Jones and Hubbell, 2006), where it was attributed to very low recruitment and high mortality rates, or in the tropical tree *Dicorynia guianensis*, linked to overlapping of generations in the adult cohort (Latouche-Hallé et al. 2003). A subsequent study of the latter species found stronger SGS in saplings (Leclerc et al., 2015), suggesting that earlier observations were probably specific to the particular study cohort. Stronger SGS in adults than in late juveniles was also found for *Prunus africana* and it was attributed to a reduction in gene flow due to past logging (Berens et al., 2014). In our study, the most probable explanation seems to be the influence of changes in contemporary management. In the managed populations of Scots pine investigated here, high felling pressure at the time of establishment of the adult cohort, together with high browsing pressure, has suppressed regeneration for decades, which is also reflected in the absence of individuals estimated between 25 and 100 years old (Fig. 2). In the last 25 years, there has been a deliberate policy to encourage regeneration in the pine forest (Mason et al., 2004), with a consequent increase in forest density. This increment in forest density appears to have maintained diversity levels, increased gene flow and produced a more randomized distribution of genotypes in the new generation.

The observed reduction in juvenile SGS shows an erosion of the structure present in the adult cohort and contemporary recovery to natural dynamics, reflecting the high capacity of the species to recover after disturbance. Overall, S_p was higher in Glen Affric than in Abernethy, as for SGS. Although the spatial extent of SGS was higher in adults at Glen Affric, S_p was slightly higher in the juvenile stage. This means more distant pairs of juveniles were less related than they would be by chance (juveniles showed a lack of relatedness among individuals at 50-80 m separation). The biological cause of this trend is not clear but, together with F_{ST} values that showed a small but significant difference among juveniles and adults, it may indicate introgression from populations not present in our sample.

4.3. Conclusions

In this study we investigated how historical and contemporary forest management have shaped patterns of genetic diversity and spatial distribution of genotypes of Scots pine. We provide evidence to show that although overall levels of genetic diversity in historically managed populations can be similar to unmanaged populations and as high as continental populations, spatial genetic structure can be considerably altered. Our results suggest that intense management practices that remove trees from the stand, such as felling, could alter fine-scale patterns of gene flow and increase genetic relatedness of individuals at fine scales with implications for inbreeding levels and, potentially, long-term adaptability. As a consequence, the extent of family clusters can be modified, as for instance in our study which increased up to 40 metres in managed sites. From a practical point of view, to ensure a broad sample of genetic variability, guidelines for seed collection should aim for minimum sampling distances between mother trees of at least 40m.

The reduction of SGS observed in juveniles following contemporary management to promote regeneration, indicates a high capacity of the species to recover after intense forest management. Here, we suggest that combining sustainable management with forest conservation practices that ensure larger effective population sizes is key to successfully maintaining genetic diversity in Scots pine. This capacity of the early stages of regeneration to capture gene flow might have implications for forest resilience and will be particularly important in the context of climate change (Alfaro et al., 2014; Fady et al., 2015; Hoffmann and Sgrò, 2011; Millar et al., 2007) under which selection pressures are expected to change.

Here we showed how investigating the spatial component of genetic diversity alongside tree demographic structure can help to detect both historical and contemporary effects of disturbances in tree populations. The effects of forest management were not reflected in overall levels of genetic diversity, but they were manifested in SGS, as has been seen in previous studies (Paffetti et al. 2012; Leclerc et al. 2015; Sjölund and Jump 2015). Therefore, incorporating a spatial

component when evaluating the effects of forest management practices is highly recommended. The evaluation of successional change is also essential to properly assess genetic dynamics within populations and to adequately detect early responses to forest management practices.

Acknowledgments

We would like to thank D. García-Martínez and E. Tordoni for help in fieldwork; R. Summers for help with selection of sampling site and for providing ring data from Abernethy; J. Sjölund and P. Ruiz-Benito for support with data analysis. This project was funded by Scottish Forestry Trust, the University of Stirling and the Centre for Ecology and Hydrology. Sampling and collection in Siberia was supported by INTERACT (grant agreement No262693) under the European Union's Seventh Framework Programme.

References

- Adams, W.T., Zuo, J., Shimizu, J.Y., Tappeiner, J.C., 1998. Impact of alternative regeneration methods on genetic diversity in coastal Douglas-fir. *For. Sci.* 44, 390–396.
- Alfaro, R.I., Fady, B., Vendramin, G.G., Dawson, I.K., Fleming, R.A., Sáenz-Romero, C., Lindig-Cisneros, R.A., Murdock, T., Vinceti, B., Navarro, C.M., Skrøppa, T., Baldinelli, G., El-Kassaby, Y.A., Loo, J., 2014. The role of forest genetic resources in responding to biotic and abiotic factors in the context of anthropogenic climate change. *For. Ecol. Manage.* 333, 76–87.
- Amos, W., Hoffman, J.I., Frodsham, A., Zhang, L., Best, S., Hill, A.V.S., 2007. Automated binning of microsatellite alleles: Problems and solutions. *Mol. Ecol. Notes* 7, 10–14.
- Auckland L.D., Bui T., Zhou Y., Shepard M., Williams C.G., 2002. *Conifer microsatellite handbook*. Corporate Press, Raleigh, N.C.
- Bain C., 2013. *The Ancient pinewoods of Scotland. A traveller's guide*. Sandstone press Ltd., Scotland.

557 Beaumont, D., Dugan, D., Evans, G., Taylor, S., 1995. Deer management and tree regeneration
558 in the RSPB reserve at Abernethy forest. *Scott. For.* 49, 155-161.

559 Belletti, P., Ferrazzini, D., Piotti, A., Monteleone, I., Ducci, F., 2012. Genetic variation and
560 divergence in Scots pine (*Pinus sylvestris* L.) within its natural range in Italy. *Eur. J. For.*
561 *Res.* 131, 1127–1138.

562 Bennett, K.D., 1984. The post-glacial history of *Pinus sylvestris* in the British Isles. *Quaternary*
563 *Sci. Rev.* 3, 133–155.

564 Berens, D.G., Braun, C., González-Martínez, S.C., Griebeler, E.M., Nathan, R., Böhning-Gaese,
565 K., 2014. Fine-scale spatial genetic dynamics over the life cycle of the tropical tree *Prunus*
566 *africana*. *Heredity (Edinb.)* 113, 401–407.

567 Bradshaw, R.H., 2004. Past anthropogenic influence on European forests and some possible
568 genetic consequences. *For. Ecol. Manage.* 197, 203–212.

569 Carlisle, A., Brown, A.H.F., 1968. *Pinus sylvestris* L. *J. Ecol.* 56, 269–307.

570 Castro, J., 1999. Seed mass versus seedling performance in Scots pine: a maternally dependent
571 trait. *New Phytol.* 144, 153–161.

572 Cavers, S., Cottrell, J.E., 2014. The basis of resilience in forest tree species and its use in
573 adaptive forest management in Britain. *Forestry* 88, 13–26.

574 Cavers, S., Degen, B., Caron, H., Lemes, M.R., Margis, R., Salgueiro, F., Lowe, A.J., 2005.
575 Optimal sampling strategy for estimation of spatial genetic structure in tree populations.
576 *Heredity (Edinb.)* 95, 281–289.

577 Chagné, D., Chaumeil, P., Ramboer, A., Collada, C., Guevara, A., Cervera, M.T., Vendramin,
578 G.G., Garcia, V., Frigerio, J.M., Echt, C., Richardson, T., Plomion, C., 2004. Cross-
579 species transferability and mapping of genomic and cDNA SSRs in pines. *Theor. Appl.*
580 *Genet.* 109, 1204–1214.

581 Chapuis, M.P., Estoup, A., 2007. Microsatellite null alleles and estimation of population
582 differentiation. *Mol. Biol. Evol.* 24, 621–631.

583 Chybicki, I.J., Dzialuk, A., Trojankiewicz, M., Slawski, M., Burczyk, J., 2008. Spatial genetic
584 structure within two contrasting stands of Scots pine (*Pinus sylvestris* L.). *Silvae Genet.*

585 57, 193–200.

586 Cottrell, J.E., Munro, R.C., Tabbener, H.E., Milner, A.D., 2003. Comparison of fine-scale
587 genetic structure using nuclear microsatellites within two British oakwoods differing in
588 population history. *For. Ecol. Manage.* 176, 287–303.

589 Davies, G.M., Gray, A., Hamilton, A., Legg, C.J., 2008. The future of fire management in the
590 British uplands. *Int. J. Biodivers. Sci. Manag.* 4, 127–147.

591 De-Lucas, A.I., González-Martínez, S.C., Vendramin, G.G., Hidalgo, E., Heuertz, M., 2009.
592 Spatial genetic structure in continuous and fragmented populations of *Pinus pinaster*
593 Aiton. *Mol. Ecol.* 18, 4564–4576.

594 DeSalle, R., Amato, G., 2004. The expansion of conservation genetics. *Nat. Rev. Genet.* 5, 702–
595 712.

596 Excoffier, L., Lischer, H.E.L., 2010. Arlequin suite ver 3.5: a new series of programs to perform
597 population genetics analyses under Linux and Windows. *Mol. Ecol. Resour.* 10, 564–567.

598 Fady, B., Cottrell, J., Ackzell, L., Alía, R., Muys, B., Prada, A., González-Martínez, S.C., 2016.
599 Forests and global change: what can genetics contribute to the major forest management
600 and policy challenges of the twenty-first century? *Reg. Environ. Chang.* 16, 927–939.

601 FAO, 2015. Global forest resources assessment 2015: How are the world’s forest changing?
602 Food and agriculture organization of the united nations, Rome, Italy.

603 FAO, 2014. The state of the world’s forest genetic resources. Commission on genetic resources
604 for food and agriculture, Rome, Italy.

605 Forest Europe, 2015. State of Europe’s forest 2015.

606 Forestry Commission, 2016. The UK forestry standard. The governments’ approach to
607 sustainable forestry. Forestry Commission, Edinburgh.

608 Forrest, G.I., 1982. Relationship of some european Scots pine populations to native Scottish
609 woodlands based on monoterpene analysis. *Forestry* 55, 19–37.

610 Forrest, G.I., 1980. Genotypic variation among native Scots pine populations in Scotland based
611 on monoterpene analysis. *Forestry* 53, 101–128.

612 García-Gil, M.R., Floran, V., Östlund, L., Mullin, T.J., Gull, B.A., 2015. Genetic diversity and

613 inbreeding in natural and managed populations of Scots pine. *Tree Genet. Genomes* 11,
 614 28.

615 Gerlach, G., Jueterbock, A., Kraemer, P., Deppermann, J., Harmand, P., 2010. Calculations of
 616 population differentiation based on GST and D: Forget GST but not all of statistics. *Mol.*
 617 *Ecol.* 19, 3845–3852.

618 González-Martínez, C., Gerber, S., Cervera, T., Martínez-Zapater, M., Gil, L., Alía, R., 2002.
 619 Seed gene flow and fine-scale structure in a Mediterranean pine (*Pinus pinaster* Ait.) using
 620 nuclear microsatellite markers. *Theor. Appl. Genet.* 104, 1290–1297.

621 Goudet, J., 1995. Computer Note. *J. Hered.* 86, 485–486.

622 Hampe, A., El Masri, L., Petit, R.J., 2010. Origin of spatial genetic structure in an expanding
 623 oak population. *Mol. Ecol.* 19, 459–471.

624 Hardesty, B.D., Dick, C.W., Kremer, A., Hubbell, S., Bermingham, E., 2005. Spatial genetic
 625 structure of *Simarouba amara* Aubl. (Simaroubaceae), a dioecious, animal-dispersed
 626 Neotropical tree, on Barro Colorado Island, Panama. *Heredity (Edinb.)* 95, 290–297.

627 Hardy, O.J., Vekemans, X., 2002. Spagedi: a versatile computer program to analyse spatial
 628 genetic structure at the individual or population levels. *Mol. Ecol. Notes* 618–620.

629 Hobbs, R., 2009. Woodland restoration in Scotland: Ecology, history, culture, economics,
 630 politics and change. *J. Environ. Manage.* 90, 2857–2865.

631 Hoffmann, A.A., Sgrò, C.M., 2011. Climate change and evolutionary adaptation. *Nature* 470,
 632 479–485.

633 Jones, F.A., Hamrick, J.L., Peterson, C.J., Squiers, E.R., 2006. Inferring colonization history
 634 from analyses of spatial genetic structure within populations of *Pinus strobus* and *Quercus*
 635 *rubra*. *Mol. Ecol.* 15, 851–861.

636 Jones, F.A., Hubbell, S.P., 2006. Demographic spatial genetic structure of the Neotropical tree,
 637 *Jacaranda copaia*. *Mol. Ecol.* 15, 3205–3217.

638 Jost, L., 2008. GST and its relatives do not measure differentiation. *Mol. Ecol.* 17, 4015–4026.

639 Jump, A.S., Peñuelas, J., 2007. Extensive spatial genetic structure revealed by AFLP but not
 640 SSR molecular markers in the wind-pollinated tree, *Fagus sylvatica*. *Mol. Ecol.* 16, 925–

641 936.

642 Jump, A.S., Rico, L., Coll, M., Peñuelas, J., 2012. Wide variation in spatial genetic structure
643 between natural populations of the European beech (*Fagus sylvatica*) and its implications
644 for SGS comparability. *Heredity* (Edinb). 108, 633–639.

645 Kettle, C.J., Hollingsworth, P.M., Jaffré, T., Moran, B., Ennos, R.A., 2007. Identifying the early
646 genetic consequences of habitat degradation in a highly threatened tropical conifer,
647 *Araucaria nemorosa* Laubenfels. *Mol. Ecol.* 16, 3581–3591.

648 Kinloch, B., Westfall, R.D., Forrest, G.I., 1986. Caledonian Scots pine: origins and genetic
649 structure. *New Phytol.* 104, 703–729.

650 Latouche-Hallé, C., Ramboer, A., Bandou, E., Caron, H., Kremer, A., 2003. Nuclear and
651 chloroplast genetic structure indicate fine-scale spatial dynamics in a neotropical tree
652 population. *Heredity* (Edinb). 91, 181–190.

653 Leclerc, T., Vimal, R., Troispoux, V., Périgon, S., Scotti, I., 2015. Life after disturbance (I):
654 changes in the spatial genetic structure of *Jacaranda copaia* (Aubl.) D. Don
655 (Bignoniaceae) after logging in an intensively studied plot in French Guiana. *Ann. For.*
656 *Sci.* 509–516.

657 Loiselle, B.A., Sork, V.L., Nason, J., Graham, C., 1995. Spatial genetic structure of a tropical
658 understory shrub, *Psychotria officinales* (Rubiaceae). *Am. J. Bot.* 82, 1420–1425.

659 Lowe, A.J., Boshier, D., Ward, M., Bacles, C.F.E., Navarro, C., 2005. Genetic resource impacts
660 of habitat loss and degradation; reconciling empirical evidence and predicted theory for
661 neotropical trees. *Heredity* (Edinb). 95, 255–273.

662 Marquardt, P.E., Epperson, B.K., 2004. Spatial and population genetic structure of
663 microsatellites in white pine. *Mol. Ecol.* 13, 3305–3315.

664 Mason, W.L., Hampson, A. and Edwards, C., 2004 Managing the pinewoods of Scotland.
665 Forestry Commission, Edinburgh.

666 Mátyás, C., Ackzell, L., Samuel, C.J.A., 2004. EUFORGEN Technical guidelines for genetic
667 conservation and use for Scots pine (*Pinus sylvestris*). International Plant Genetic
668 Resources Institute, Rome, Italy.

669 McVean, D.N., 1963. Ecology of Scots pine in the Scottish Highlands. *J. Ecol.* 51, 671–686.

670 Millar, C.I., Stephenson, N.L., Stephens, S.L., 2007. Climate change and forest of the future:
671 managing in the face of uncertainty. *Ecol. Appl.* 17, 2145–2151.

672 Naydenov, K.D., Naydenov, M.K., Tremblay, F., Alexandrov, A., Aubin-Fournier, L.D., 2011.
673 Patterns of genetic diversity that result from bottlenecks in Scots pine and the implications
674 for local genetic conservation and management practices in Bulgaria. *New For.* 42, 179–
675 193.

676 Naydenov, K.D., Senneville, S., Beaulieu, J., Tremblay, F., Bousquet, J., 2007. Glacial
677 vicariance in Eurasia: mitochondrial DNA evidence from Scots pine for a complex
678 heritage involving genetically distinct refugia at mid-northern latitudes and in Asia Minor.
679 *BMC Evol. Biol.* 7, 233.

680 Ng, K.K.S., Lee, S.L., Koh, C.L., 2004. Spatial structure and genetic diversity of two tropical
681 tree species with contrasting breeding systems and different ploidy levels. *Mol. Ecol.* 13,
682 657–669.

683 Nowakowska, J.A., Zachara, T., Konecka, A., 2014. Genetic variability of Scots pine (*Pinus*
684 *sylvestris* L.) and Norway spruce (*Picea abies* L. Karst.) natural regeneration compared
685 with their maternal stands. *For. Res. Pap.* 75, 47–54.

686 O’Sullivan, P.E., 1973. Land-use changes in the forest of Abernethy, Inverness-shire, 1750 -
687 1900. *Scott. Geogr. Mag.* 89, 95–106.

688 Ortego, J., Bonal, R., Muñoz, A., 2010. Genetic consequences of habitat fragmentation in long-
689 lived tree species: the case of the mediterranean holm oak (*Quercus ilex*, L.). *J. Hered.*
690 101, 717–726.

691 Paffetti, D., Travaglini, D., Buonamici, A., Nocentini, S., Vendramin, G.G., Giannini, R.,
692 Vettori, C., 2012. The influence of forest management on beech (*Fagus sylvatica* L.) stand
693 structure and genetic diversity. *For. Ecol. Manage.* 284, 34–44.

694 Pandey, M., Gailing, O., Hattemer, H.H., Finkeldey, R., 2012. Fine-scale spatial genetic
695 structure of sycamore maple (*Acer pseudoplatanus* L.). *Eur. J. For. Res.* 131, 739–746.

696 Parker, K.C., Hamrick, J.L., Parker, A.J., Nason, J.D., 2001. Fine-scale genetic structure in

697 *Pinus clausa* (Pinaceae) populations: effects of disturbance history. *Heredity* (Edinb). 87,
698 99–113.

699 Piotti, A., Leonardi, S., Heuertz, M., Buiteveld, J., Geburek, T., Gerber, S., Kramer, K., Vettori,
700 C., Vendramin, G.G., 2013. Within-population genetic structure in beech (*Fagus sylvatica*
701 L.) Stands characterized by different disturbance histories: does forest management
702 simplify population substructure? *PLoS One* 8, e73391.

703 Provan, J., Soranzo, N., Wilson, N.J., McNicol, J.W., Forrest, G.I., Cottrell, J.E., Powell, W.,
704 1998. Gene-pool variation in caledonian and European Scots pine (*Pinus sylvestris* L.)
705 revealed by chloroplast simple-sequence repeats. *Proc. Biol. Sci.* 265, 1697–1705.

706 Prus-Glowacki, W., Stephan, B.R., 1994. Genetic variation of *Pinus sylvestris* from Spain in
707 relation to other European populations. *Silvae Genet.* 43, 7–14.

708 Prus-Glowacki, W., Urbaniak, L., Bujas, E., Curtu, A.L., 2012. Genetic variation of isolated and
709 peripheral populations of *Pinus sylvestris* (L.) from glacial refugia. *Flora - Morphol.*
710 *Distrib. Funct. Ecol. Plants* 207, 150–158.

711 R Core Team, 2013. R: A language and environment for statistical computing. R Foundation for
712 Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.

713 Rajora, O.P., Pluhar, S.A., 2003. Genetic diversity impacts of forest fires, forest harvesting, and
714 alternative reforestation practices in black spruce (*Picea mariana*). *Theor. Appl. Genet.*
715 106, 1203–1212.

716 Rajora, O.P., Rahman, M.H., Buchert, G.P., Dancik, B.P., 2000. Microsatellite DNA analysis of
717 genetic effects of harvesting in old-growth eastern white pine (*Pinus strobus*) in Ontario,
718 Canada. *Mol. Ecol.* 9, 339–348.

719 Robledo-Arnuncio, J.J., 2011. Wind pollination over mesoscale distances: an investigation with
720 Scots pine. *New Phytol.* 190, 222–233.

721 Robledo-Arnuncio, J.J., Alía, R., Gil, L., 2004a. Increased selfing and correlated paternity in a
722 small population of a predominantly outcrossing conifer, *Pinus sylvestris*. *Mol. Ecol.* 13,
723 2567–2577.

724 Robledo-Arnuncio, J.J., Smouse, P.E., Gil, L., Alía, R., 2004b. Pollen movement under

725 alternative silvicultural practices in native populations of Scots pine (*Pinus sylvestris* L.) in
 726 central Spain. *For. Ecol. Manage.* 197, 245–255.

727 Rozas, V., Zas, R., Solla, A., 2009. Spatial structure of deciduous forest stands with contrasting
 728 human influence in northwest Spain. *Eur. J. For. Res.* 128, 273–285.

729 Scalfi, M., Piotti, A., Rossi, M., Piovani, P., 2009. Genetic variability of Italian southern Scots
 730 pine (*Pinus sylvestris* L.) populations: the rear edge of the range. *Eur. J. For. Res.* 128,
 731 377–386.

732 Schaberg, P.G., DeHayes, D.H., Hawley, G.J., Nijensohn, S.E., 2008. Anthropogenic alterations
 733 of genetic diversity within tree populations: Implications for forest ecosystem resilience.
 734 *For. Ecol. Manage.* 256, 855–862.

735 Sebastiani, F., Pinzauti, F., Kujala, S.T., González-Martínez, S.C., Vendramin, G.G., 2011.
 736 Novel polymorphic nuclear microsatellite markers for *Pinus sylvestris* L. *Conserv. Genet.*
 737 *Resour.* 4, 231–234.

738 Shaw, H., 2006. Recent pine woodland dynamics in east Glen Affric, northern Scotland, from
 739 highly resolved palaeoecological analyses. *Forestry* 79, 331–340.

740 Sinclair, W.T., Morman, J.D., Ennos, R.A., 1998. Multiple origins for Scots pine (*Pinus*
 741 *sylvestris* L.) in Scotland: evidence from mitochondrial DNA variation. *Heredity* (Edinb).
 742 80, 233–240.

743 Sjölund, M.J., Jump, A.S., 2015. Coppice management of forests impacts spatial genetic
 744 structure but not genetic diversity in European beech (*Fagus sylvatica* L.). *For. Ecol.*
 745 *Manage.* 336, 65–71.

746 Smouse, P.E., Peakall, R.O.D., 1999. Spatial autocorrelation analysis of individual multiallele
 747 and multilocus genetic structure 82, 561–573.

748 Smout, T. C., 2003. *People and Woods in Scotland: a History*, Edinburgh University Press,
 749 Edinburgh.

750 Smout, T. C., MacDonald, A.R., Watson, F., 2005. *A history of the native woodland of Scotland*
 751 *1500-1920*, Edinburgh University Press, Edinburgh

752 Soranzo, N., Provan, J., Powell, W., 1998. Characterization of microsatellite loci in *Pinus*

753 *sylvestris* L. Mol. Ecol. 7, 1260–1261.

754 Soto, A., Lorenzo, Z., Gil, L., 2007. Differences in fine-scale genetic structure and dispersal in

755 *Quercus ilex* L. and *Q. suber* L.: consequences for regeneration of mediterranean open

756 woods. Heredity (Edinb). 99, 601–607.

757 Steven, H.M., Carlisle, A., 1959. The native pinewoods of Scotland. GC Book Publishers,

758 Edinburgh.

759 Summers, R.W., Wilkinson, N.I., Wilson, E.R., 2008. Age structure and history of stand types

760 of *Pinus sylvestris* in Abernethy Forest, Scotland. Scand. J. For. Res. 23, 28–37.

761 Szwagrzyk, J., Czerwczak, M., 1993. Spatial patterns of trees in natural forests of East-Central

762 Europe. J. Veg. Sci. 4, 469–476.

763 Tipping, R., Ashmore, P., Davies, A.L., Haggart, B.A., Moir, A., Newton, A., Sands, R.,

764 Skinner, T., Tisdall, E., 2008. Prehistoric *Pinus* woodland dynamics in an upland

765 landscape in northern Scotland: the roles of climate change and human impact. Veg. Hist.

766 Archaeobot. 17, 251–267.

767 Troupin, D., Nathan, R., Vendramin, G.G., 2006. Analysis of spatial genetic structure in an

768 expanding *Pinus halepensis* population reveals development of fine-scale genetic

769 clustering over time. Mol. Ecol. 15, 3617–3630.

770 Vander Wall, S.B., 2003. Effects of seed size of wind-dispersed pines (*Pinus*) on secondary

771 seed dispersal and the caching behavior of rodents. Oikos 100, 25–34.

772 Van Oosterhout, C., Hutchinson, W.F., Wills, D.P.M., Shipley, P., 2004. Micro-Checker:

773 software for identifying and correcting genotyping errors in microsatellite data. Mol. Ecol.

774 Notes 4, 535–538.

775 Vekemans, X., Hardy, O.J., 2004. New insights from fine-scale spatial genetic structure

776 analyses in plant populations. Mol. Ecol. 13, 921–935.

777 Wachowiak, W., Iason, G.R., Cavers, S., 2013. Among population differentiation at nuclear

778 genes in native Scots pine (*Pinus sylvestris* L.) in Scotland. Flora 208, 79–86.

779 Wachowiak, W., Salmela, M.J., Ennos, R.A., Iason, G., Cavers, S., 2011. High genetic diversity

780 at the extreme range edge: nucleotide variation at nuclear loci in Scots pine (*Pinus*

sylvestris L.) in Scotland. *Heredity* (Edinb). 106, 775–787.

Wachowiak, W., Wójkiewicz, B., Cavers, S., Lewandowski, A., 2014. High genetic similarity between Polish and North European Scots pine (*Pinus sylvestris* L.) populations at nuclear gene loci. *Tree Genet. Genomes* 10, 1015–1025.

Wang, X.R., Szmidt, A.E., Lindgren, D., 1991. Allozyme differentiation among populations of *Pinus sylvestris* (L.) from Sweden and China. *Hereditas* 114, 219–226.

Williams, D.A., Wang, Y., Borchetta, M., Gaines, M.S., 2007. Genetic diversity and spatial structure of a keystone species in fragmented pine rockland habitat. *Biol. Conserv.* 138, 256–268.

Yamagishi, H., Tomimatsu, H., Ohara, M., 2007. Fine-scale spatial genetic structure within continuous and fragmented populations of *Trillium camschatcense*. *J. Hered.* 98, 367–372.

Young, A., Boyle, T., Brown, T., 1996. The population genetic consequences of habitat fragmentation for plants. *Tree* 11, 413–418.

Tables

Table 1: Summary of multilocus genetic diversity and SGS estimators for each study site and life stage.

Figures

Fig. 1: Tree diameter (d.b.h.) distribution in the three study sites: Abernethy (ABE), Glen Affric (GLA) and the unmanaged site (UNM). Juvenile stem diameter was measured at 10 cm height. Data are presented in intervals of 5 cm.

Fig. 2: Relationship between d.b.h. and age for the managed site of Abernethy (ABE) and the unmanaged site (UNM). Lines of best fit are represented by solid lines and 95% CI by dashed lines. Dots represent observed values.

Fig. 3: Spatial autocorrelograms for each study site: Abernethy (ABE), Glen Affric (GLA) and the unmanaged site (UNM); and life stage (adult and juvenile) based on the kinship coefficient F_{ij} , estimated from 12 microsatellite loci, and consecutive 10 m distance classes (note that for the unmanaged stand distance classes were combined between 30 to 60 metres). Shaded areas represent 95% confident intervals obtained from 10,000 permutations of genotypes among locations. Black bars around mean F_{ij} values represent standard errors derived through jackknifing over loci.

Appendix A. Supplementary material

Table A1: Genetic diversity estimators for each locus, study site and life stage.

Table A2: Pairwise F_{ST} values (below diagonal) and differentiation index D (Jost, 2008) (above diagonal) among study sites and life stages.

Table 1: Summary of multilocus genetic diversity and SGS estimators for each study site and life stage.

Study site	Life stage	N	Genetic diversity estimators				Spatial genetic structure estimators			
			A	A_R	H_E	F_{IS}	$F_{(I)}$	SGS_{MAX} (m)	$b_F \pm SE$	$Sp \pm SE$
Abernethy	Adult	181	9.50	7.11	0.587	0.052**	0.0291***	20	-0.0044 \pm 0.0006***	0.0045 \pm 0.0028
	Juvenile	170	9.25	6.72	0.583	0.080**	0.0183***	18	-0.0028 \pm 0.0009**	0.0029 \pm 0.0023
Glen Affric	Adult	165	8.92	6.79	0.568	0.063**	0.0298***	40	-0.0097 \pm 0.0023***	0.0098 \pm 0.0010
	Juvenile	131	9.25	6.74	0.561	0.049**	0.0156***	20	-0.0118 \pm 0.0027***	0.0119 \pm 0.0006
Unmanaged	Adult	57	7.58	6.51	0.576	0.012	-0.0033	0	0.0006 \pm 0.0005	-0.0006 \pm 0.0005
	Juvenile	73	8.17	6.94	0.582	0.021	0.0067	5	-0.0017 \pm 0.0010*	0.0018 \pm 0.0011

N , sample size; A , mean number of alleles per locus; A_R , rarefied allelic richness; H_E , expected heterozygosity; F_{IS} , inbreeding coefficient. $F_{(I)}$, Kinship coefficient for first distance class (0-10m); SGS_{MAX} , greatest distance at which the Kinship coefficient of a given distance class $F(d)$ is significant at $p < 0.05$; $b_F \pm SE$, regression slope of the Kinship coefficient F_{ij} computed among all individuals against geographical distances \pm standard error; $Sp \pm SE$, Sp statistic \pm standard error. Significant P -values are indicated as * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. P -values for F_{IS} are obtained after 10,000 permutations of gene copies within individuals of each stand.

Fig. 1: Tree diameter (d.b.h.) distribution in the three study sites: Abernethy (ABE), Glen Affric (GLA) and the unmanaged site (UNM). Juvenile stem diameter was measured at 10 cm height. Data are presented in intervals of 5 cm.

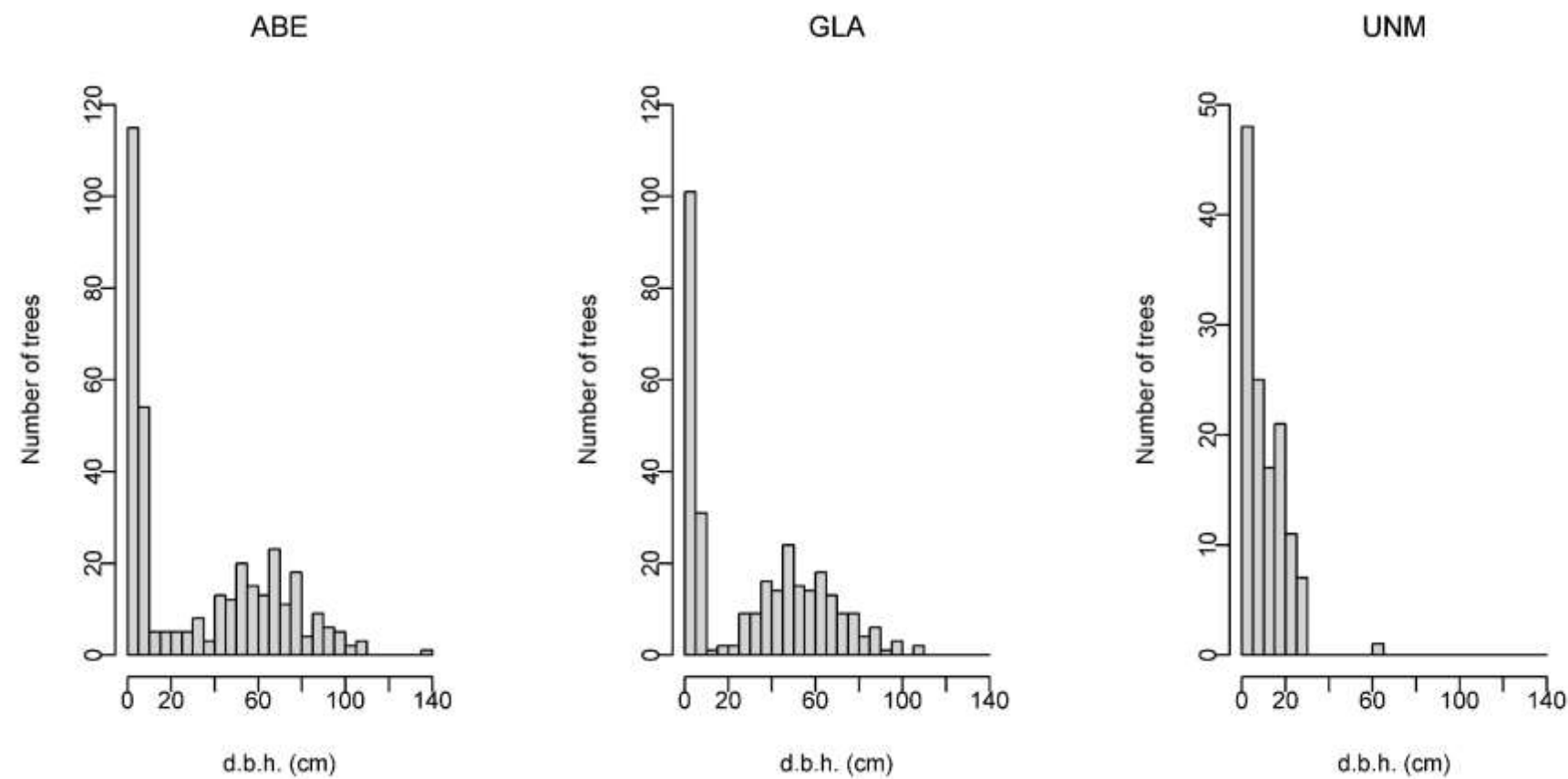


Fig. 2: Relationship between d.b.h. and age for the managed site of Abernethy (ABE) and the unmanaged site (UNM). Lines of best fit are represented by solid lines and 95% CI by dashed lines. Dots represent observed values.

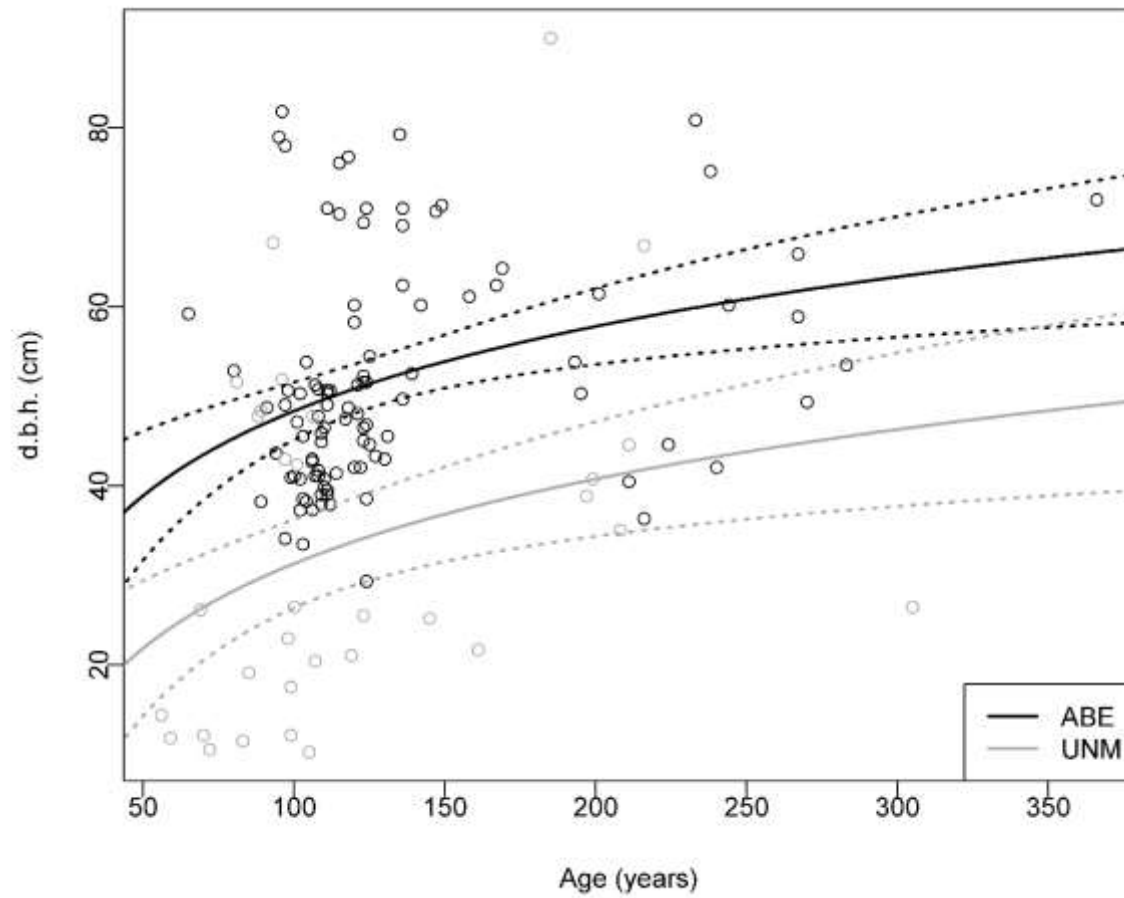
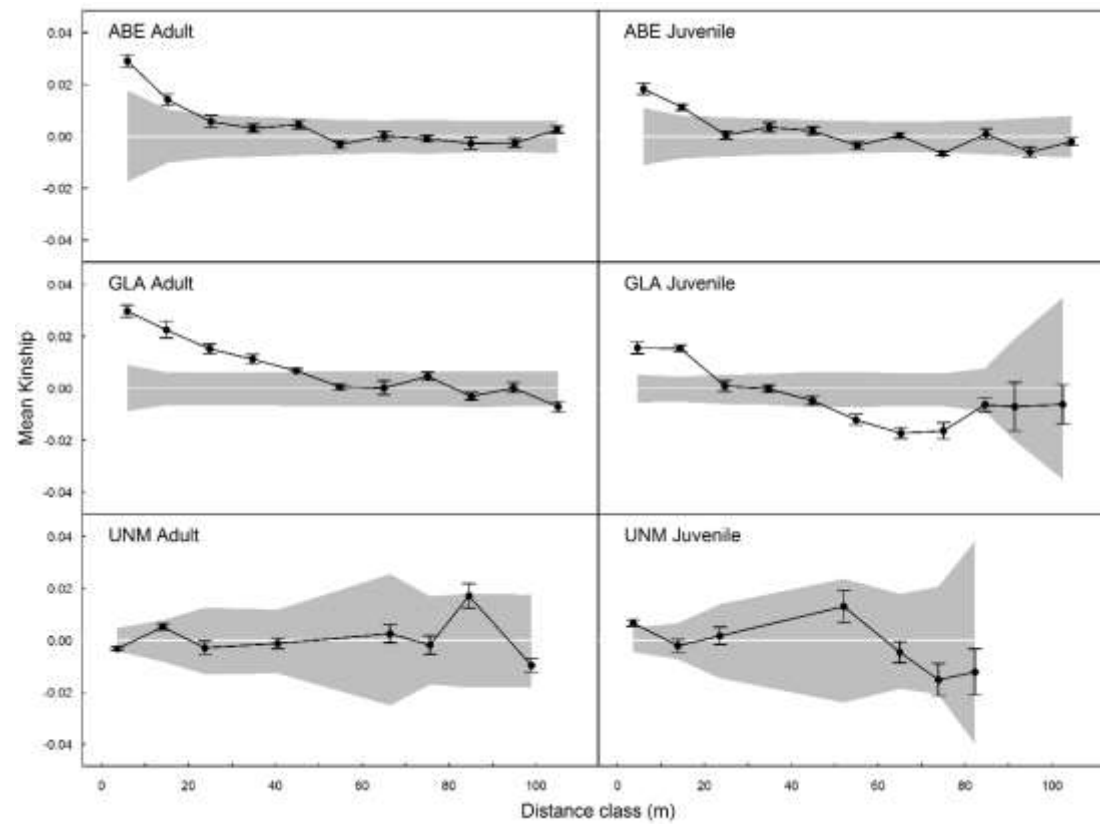


Fig. 3: Spatial autocorrelograms for each study site: Abernethy (ABE), Glen Affric (GLA) and the unmanaged site (UNM); and life stage (adult and juvenile) based on the kinship coefficient F_{ij} , estimated from 12 microsatellite loci, and consecutive 10 m distance classes (note that for the unmanaged stand distance classes were combined between 30 to 60 metres). Shaded areas represent 95% confident intervals obtained from 10,000 permutations of genotypes among locations. Black bars around mean F_{ij} values represent standard errors derived through jackknifing over loci.



Appendix A. Supplementary material

Table A1: Genetic diversity estimators for each locus, study site and life stage.

Locus	Life stage	Abernethy					Glen Affric					Unmanaged				
		<i>N</i>	<i>A</i>	<i>A_R</i>	<i>H_E</i>	<i>F_{IS}</i>	<i>N</i>	<i>A</i>	<i>A_R</i>	<i>H_E</i>	<i>F_{IS}</i>	<i>N</i>	<i>A</i>	<i>A_R</i>	<i>H_E</i>	<i>F_{IS}</i>
PtTX4001	Adult	181	11	9.28	0.8306	-0.028	165	9	7.59	0.7783	0.03	57	7	6.22	0.5951	-0.002
	Juvenile	170	12	9.32	0.8430	-0.06	131	11	8.79	0.8074	0.054	73	9	6.22	0.5073	0.028
PtTX4011	Adult	181	7	4.61	0.5920	0.099*	165	7	5.12	0.5423	0.213***	57	6	5.66	0.6717	0.204*
	Juvenile	170	6	4.73	0.6144	0.22***	131	6	5.05	0.6094	0.097	73	5	4.96	0.6922	0.3*
psy144	Adult	181	5	3.08	0.1166	-0.042	165	5	3.12	0.1380	-0.054	57	2	1.88	0.0517	-0.018
	Juvenile	170	5	2.88	0.0804	-0.024	131	5	3.2	0.1581	-0.067	73	3	2.39	0.1293	-0.06
psy117	Adult	181	8	6.32	0.7820	0.054	165	10	6.97	0.7907	-0.004	57	8	7.03	0.8224	-0.065
	Juvenile	170	8	5.98	0.7600	0.133**	131	8	6.56	0.7580	0.016	73	7	6.8	0.8247	-0.025
psy142	Adult	181	5	4.15	0.6466	0	165	6	5.22	0.6669	0.019	57	4	3.51	0.6479	-0.084
	Juvenile	170	6	4.34	0.6632	0.104*	131	6	5.07	0.6551	0.01	73	5	4.32	0.6411	-0.155*
psy12	Adult	181	3	2.17	0.3193	0.163*	165	3	2.18	0.2727	-0.096	57	2	2	0.3354	0.059
	Juvenile	170	3	2.17	0.3539	0.087	131	3	2.23	0.2386	0.393***	73	2	2	0.2314	-0.017
psy116	Adult	181	7	5.95	0.7862	-0.03	165	6	5.5	0.7736	0.011	57	6	5.5	0.7399	-0.092
	Juvenile	170	8	5.95	0.7720	0.063	131	7	5.42	0.7512	-0.024	73	6	5.87	0.7598	-0.01
psy157	Adult	181	5	4.23	0.3652	0.002	165	6	4.52	0.3483	-0.009	57	4	3.99	0.3892	-0.128
	Juvenile	170	5	4.19	0.3517	0.064	131	5	4.05	0.2984	-0.024	73	5	4.39	0.5168	-0.087
CTG4698	Adult	181	8	6.24	0.6044	0.019	165	8	5.17	0.5635	-0.043	57	5	5	0.6500	0.049
	Juvenile	170	6	5.34	0.6124	-0.034	131	6	5.27	0.5721	-0.068	73	5	4.64	0.6065	-0.016
SPAC7.14	Adult	181	29	19.08	0.9174	0.194***	165	26	18.6	0.9150	0.236***	57	22	17.95	0.9023	0.09*
	Juvenile	170	28	17.13	0.9093	0.179***	131	28	17.83	0.9072	0.21***	73	28	22.47	0.9513	0.097**

SPAC12.5	Adult	181	21	16.15	0.8989	-0.007	165	17	14.62	0.9058	0.098***	57	22	16.58	0.8475	0.048
	Juvenile	170	19	15.33	0.8956	0.054*	131	22	14.58	0.8814	0.005	73	19	15.85	0.8629	0.032
psy136	Adult	181	5	4.06	0.1877	0.438***	165	4	2.82	0.1166	0.216***	57	3	2.76	0.2607	-0.01
	Juvenile	170	5	3.23	0.1451	0.108	131	4	2.82	0.0897	-0.029	73	4	3.35	0.2578	-0.01

N , sample size; A , mean number of alleles per locus; A_R , rarefied allelic richness; H_E , expected heterozygosity; F_{IS} , inbreeding coefficient. Significant P -values are indicated as

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. P -values for F_{IS} are obtained after 10,000 permutations of gene copies within individuals of each stand.

Table A2: Pairwise F_{ST} values (below diagonal) and differentiation index D (Jost, 2008) (above diagonal) among study sites and life stages.

	ABE Adults	ABE Juveniles	GLA Adults	GLA Juveniles	UNM Adults	UNM Juveniles
ABE Adults	-	-0.00134	0.01367***	0.01694***	0.09089***	0.08407***
ABE Juveniles	-0.00085	-	0.01925***	0.01836***	0.09777***	0.09615***
GLA Adults	0.00531***	0.00504***	-	0.01223**	0.08486***	0.08469***
GLA Juveniles	0.00794***	0.00712***	0.00514***	-	0.09852***	0.09642***
UNM Adults	0.04973***	0.05174***	0.04434***	0.05228***	-	0.00843
UNM Juveniles	0.04923***	0.05132***	0.04586***	0.05382***	-0.00254	-

ABE refers to Abernethy, GLA refers to Glen Affric, UNM refers to the unmanaged site. Significant P -values are indicated as * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.