

**Molecular Cloning and Functional Characterization of Fatty Acyl Desaturase and Elongase cDNAs Involved in the Production of Eicosapentaenoic and Docosahexaenoic Acids from  $\alpha$ -Linolenic Acid in Atlantic Salmon (*Salmo salar*)**

Nicola Hastings, Morris K. Agaba, Douglas R. Tocher\*, Xiaozhong Zheng, Cathryn A. Dickson, James R. Dick and Alan J. Teale

*Institute of Aquaculture, University of Stirling, Stirling FK9 4LA, Scotland, United Kingdom*

\*Corresponding author: telephone +44 1786 467996; fax +44 1786 472133; e-mail: [d.r.tocher@stir.ac.uk](mailto:d.r.tocher@stir.ac.uk)

**Abstract:** Fish are the only major dietary source for humans of omega-3 highly unsaturated fatty acids (HUFA) and, with declining fisheries, farmed fish such as Atlantic salmon (*Salmo salar*) constitute an increasing proportion of the fish in the human diet. However, the current high use of fish oils, derived from wild capture marine fisheries, in aquaculture feeds is not sustainable in the longer term, and will constrain continuing growth of aquaculture activities. A greater understanding of how fish metabolise and biosynthesise HUFA may lead to effective use of more sustainable aquaculture diets. The study described here contributes to an effort to determine the molecular genetics of the HUFA biosynthetic pathway in salmon, with the overall aim being to determine mechanisms for optimising the use of vegetable oils in Atlantic salmon culture. In this paper we describe the cloning and functional characterisation of two genes from salmon involved in the biosynthesis of HUFA. A salmon desaturase cDNA, SalDes, was isolated that included an open reading frame (ORF) of 1362 bp specifying a protein of 454 amino acids. The protein sequence included all the characteristic features of microsomal fatty acid desaturases, including three histidine boxes, two transmembrane regions, and an N-terminal cytochrome b<sub>5</sub> domain containing a haem-binding motif similar to that of other fatty acid desaturases. Functional expression in the yeast, *Saccharomyces cerevisiae*, showed SalDes is predominantly an omega-3  $\Delta^5$  desaturase, a key enzyme in the synthesis of eicosapentaenoic acid (20:5n-3) from  $\alpha$ -linolenic acid (18:3n-3). The desaturase showed only low levels of  $\Delta^6$  activity towards C<sub>18</sub> polyunsaturated fatty acids. In addition, a fatty acid elongase cDNA, SalElo, was isolated that includes an ORF of 888 bp, specifying a protein of 295 amino acids. The protein sequence of SalElo includes characteristic features of microsomal fatty acid elongases, including a histidine box and a transmembrane region. Upon expression in yeast, SalElo showed broad substrate specificity for polyunsaturated fatty acids with a range of chain lengths, with the rank order being C<sub>18</sub> > C<sub>20</sub> > C<sub>22</sub>. Thus, all fatty acid elongase activities required for the biosynthesis of docosahexaenoic acid (22:6n-3) from 18:3n-3 are displayed by this one polypeptide product.

**Key words:** Atlantic salmon, highly unsaturated fatty acids, desaturase, elongase.

## INTRODUCTION

Generally declining catches from wild fisheries have resulted in an increasing proportion of fish for human consumption being provided by aquaculture, which is currently expanding at over 10% per annum (Tidwell and Allan, 2002). However, much aquaculture, including salmonid (salmon and trout) culture, is itself dependent upon wild capture fisheries for the provision of fish meals and oils that have traditionally been the predominant protein and lipid sources (Sargent and Tacon, 1999). This was a sound strategy as both are readily accepted and digested by fish and, until now, were readily available and inexpensive (Wilson, 1989; Sargent et al., 1989; Barlow, 2000). However, stagnation in industrial fisheries, along with the increased demand for fish oils, has dictated that alternatives to fish oil must be found if aquaculture is to continue to expand and supply more of the global demand for fish (Barlow, 2000).

The only sustainable alternative to fish oils are plant (vegetable) oils, some of which are rich in C<sub>18</sub> polyunsaturated fatty acids (PUFA) such as 18:2n-6 and 18:3n-3. However, vegetable oils are devoid of the n-3 highly unsaturated fatty acids (HUFA), eicosapentaenoic (EPA; 20:5n-3) and docosahexaenoic (DHA; 22:6n-3) acids, that are abundant in fish oils (Sargent et al., 2002). The extent to which animals, including fish, can convert C<sub>18</sub> PUFA to C<sub>20/22</sub> HUFA varies with species, and is associated with their capacity for microsomal fatty acyl desaturation and elongation. Salmonid fish, including Atlantic salmon (*Salmo salar*), are capable of producing DHA from 18:3n-3 (Buzzi et al., 1996; Tocher et al., 1997; Bell et al., 2001a) and so express all the desaturase and elongase activities necessary for this biosynthetic pathway (Sargent et al., 2002). Salmon fed diets containing vegetable oils show significantly increased activity of the fatty acyl desaturation/elongation pathway compared to fish fed standard diets containing fish oil (Bell et al. 1997, 2001b, 2002; Tocher et al., 1997, 2000, 2001). Nonetheless, tissue fatty acid compositions in salmon fed vegetable oils are characterised by increased levels of C<sub>18</sub> PUFA and decreased levels of C<sub>20/22</sub> HUFA, compromising their nutritional value to the human consumer (Bell et al., 2001b, 2002). Even in salmon fed linseed oil containing high levels of 18:3n-3, the increased activity of the desaturation/elongation pathway cannot convert 18:3n-3 to EPA and DHA efficiently enough to prevent the accumulation of 18:3n-3 and depletion of EPA and DHA in the tissues (Bell et al., 1997; Tocher et al., 2000, 2001).

An understanding of the molecular basis of HUFA biosynthesis would underpin efforts to address this problem. However, until recently, although the biochemical pathways involved in PUFA synthesis were described, little was known of the enzymes involved and of the factors affecting their function(s). Significant progress has now been made in characterizing the desaturases and elongases involved in HUFA synthesis (Tocher et al., 1998). Full-length cDNAs for  $\Delta 6$  desaturases have been isolated from the filamentous fungi *Mortierella alpina* (Huang et al., 1999),

the nematode *Caenorhabditis elegans* (Napier et al., 1998), rat (Aki et al., 1999), mouse and human (Cho et al., 1999a). Fatty acid  $\Delta 5$  desaturase genes have been isolated from *M. alpina* (Michaelson et al., 1998a) *C. elegans* (Michaelson et al., 1998b; Watts and Browse, 1999) and human (Cho et al., 1999b; Leonard et al., 2000a). The genes encoding putative PUFA elongase enzymes have been cloned from *M. alpina* (Parker-Barnes et al., 2000), *C. elegans* (Beaudion et al., 2000) and human (Leonard et al., 2000b). When expressed in *S. cerevisiae* all of these elongase genes were shown to encode enzymes that elongated C<sub>18</sub> PUFA to C<sub>20</sub>, and the human elongase also elongated C<sub>20</sub> PUFA to C<sub>22</sub>.

The availability of sequence data has made it possible to study the genes of the HUFA biosynthesis pathway in fish, and recently we isolated two cDNAs from zebrafish (*Danio rerio*), one with high similarity to mammalian  $\Delta 6$  desaturase genes (Hastings et al., 2001) and one with high similarity to the human elongase (Agaba et al., 2003). Heterologous expression in *S. cerevisiae* indicated that the zebrafish genes were unique in that the desaturase cDNA encoded an enzyme having both  $\Delta 6$  and  $\Delta 5$  desaturase activities and the elongase cDNA encoded an enzyme with activity towards C<sub>18</sub>, C<sub>20</sub> and C<sub>22</sub> PUFA. The objective of the present study is to determine the molecular genetics of the HUFA biosynthetic pathway in salmon with the overall aim being to determine mechanisms for optimising the use of vegetable oils in Atlantic salmon culture. As a first step, this paper describes the cloning and functional characterisation of two genes of Atlantic salmon involved in the biosynthesis of EPA and DHA.

## **MATERIALS AND METHODS**

### **Cloning of a salmon PUFA desaturase cDNA and sequence analysis.**

Desaturase sequences of fish, including the zebrafish  $\Delta 6/\Delta 5$  desaturase (GenBank accession no. AF309556) and the putative  $\Delta 6$  desaturases from rainbow trout (GenBank accession no. AF301910) and carp (GenBank accession no. AF309557), were aligned to enable the design of the degenerate primer, Fish 6for (5'-CCCAAGCTTGAGGATGGGAGTGG-3'). This was used in conjunction with T7PolyT (5'-TACGACTCACTATAGGGCGTGCAGTTTTTTTTTTT-3') for the PCR isolation of the salmon desaturase using cDNA produced from total RNA isolated from liver tissue of Atlantic salmon fed a standard pellet diet based on fish meal and oil. Briefly, cDNA was synthesized from salmon liver total RNA using Moloney murine leukaemia virus reverse transcriptase (Promega, Madison, WI, USA) primed with T7PolyT. A portion of this cDNA was then subjected to PCR amplification (Ready-to-Go PCR beads; Amersham Biosciences UK Ltd., Little Chalfont, Bucks., U.K.) with the T7PolyT primer and the degenerate Fish 6for primer as above which contains the predicted initiation codon. Amplification involved an initial denaturation

step at 95 °C for 60 s, followed by 30 cycles of denaturation at 95 °C for 30 s, annealing at 58 °C for 30 s, and extension at 72 °C for 2 min. The products were cloned into the Bluescript KSII<sup>+</sup> vector (Stratagene, La Jolla, CA, USA) using standard methods, and nucleotide sequences were determined by standard dye terminator chemistry using a Perkin Elmer ABI-377 DNA sequencer following the manufacturers protocols (Perkin Elmer, Applied Biosystems). Deduced amino acid sequences were aligned using ClustalX and sequence phylogenies were predicted using the Neighbour Joining method of Saitou and Nei (1987). Confidence in the resulting phylogenetic tree branch topology was measured by bootstrapping through 1000 iterations.

### **Cloning of a salmon PUFA elongase cDNA**

The PUFA elongase sequence (AF206662) of *M. alpina* was used to query the GenBank EST database (at NCBI) for homologues using the tblastn programme. Several vertebrate ESTs including one from zebrafish (GeneBank accession no. BF157708) were identified that had high similarity to the query sequence. Degenerate oligonucleotides were designed based on the conserved motifs (GPRD, LWYYF, EFMD SFF) identified by aligning the deduced amino acid sequences of PUFA elongases from human (ELOVL5, AF231981), *Xenopus laevis* (BG813561) and zebrafish (BF157708, AF532782). The primer sequences were UniElo1A (CCTGTGGTGGTAYTAYTT) and UniElo1AA (GARTTY ATGGACACNTTCTTCTT) and targeted the LWYYF and EFMD SFFF motifs, respectively. These primers were used in 3'RACE PCR to clone the partial transcript from Atlantic salmon liver cDNA using the SMART RACE system (Clontech Laboratories UK Ltd., Basingstoke, U.K.). PCR fragments were cloned into Bluescript KSII<sup>+</sup> vector and the sequences determined and analysed as described above. The sequence of the partial cDNA was then used to design primers for 5'RACE PCR and so clone the 5' end of the transcript.

### **Heterologous expression of desaturase and elongase ORFs in *Saccharomyces cerevisiae*.**

Functional characterisation of the genes was by expression of the open reading frames (ORFs) of the salmon putative fatty acid desaturase (SalDes) and elongase (SalElo) sequences in *Saccharomyces cerevisiae*. Expression primers were designed for amplification of the ORF's from salmon liver cDNA. These primers also carried restriction sites to enable cloning into the appropriate expression vectors. For SalDes, the sequence of the sense primer SalpYESFor (CCCAAGCTTACTATGGGGGGCGGAGGCG) contained an *HindIII* site (underlined) and the antisense primer SalPYESRev (CCGCTCGAGTCATTTATGGAGATATGCAT) contained an *XhoI* site (underlined). For SalElo, the sequence of the sense primer SalEloXA (AAGAATTCAAGCTTCTAGGGTCAGAAATGGAG) contained an *EcoRI* site (underlined), and

the antisense primer SalEloX1B (AACTCGAGACAGTATTCAAGCTTCAGTCCC) contained an *XhoI* site (underlined). PCR was performed using high fidelity DNA polymerase (Roche Diagnostics Ltd., Lewes, East Sussex, UK) following the manufacturer's instruction. Following PCR, the DNA fragments were restricted with the appropriate enzymes, *HindIII* and *XhoI* for SalDes, and *EcoRI* and *XhoI* for SalElo, and ligated into similarly digested yeast expression vectors pYES2 (Invitrogen Ltd, Paisley, UK) and pYX222 (R & D Systems Europe Ltd., Abingdon, Oxon, UK), respectively. These were then used to transform Top10F' *E. coli* competent cells (Invitrogen Ltd, Paisley, UK) which were screened for the presence of recombinant plasmids. Transformation of the yeast, *Saccharomyces cerevisiae* (strain InvSc1) with the recombinant plasmids was carried out using the S.c.EasyComp Transformation Kit (Invitrogen Ltd, Paisley, UK). Selection of yeast transformants containing the SalDes/pYES2 and SalElo/pYX222 constructs was by growth on uracil and histidine drop-out media, respectively.

Culture of the recombinant yeast SalDes/pYES2 was carried out in SCMM<sup>-uracil</sup> broth as described previously (Hastings et al., 2001), using galactose induction of gene expression. Expression of SalElo/pYX222, which was under control of a constitutive promoter, was grown on SCMM<sup>-histidine</sup> broth containing 2% glucose. Each culture was supplemented with one of the following PUFA substrates; for the SalDes cDNA,  $\alpha$ -linolenic acid (18:3n-3), linoleic acid (18:2n-6), eicosatetraenoic acid (20:4n-3), dihomo- $\gamma$ -linoleic acid (20:3n-6), docosapentaenoic acid (22:5n-3), docosatetraenoic acid (22:4n-6), tetracosapentaenoic acid (24:5n-3) and tetracosatetraenoic acid (24:4n-6); and for the SalElo cDNA, stearidonic acid (18:4n-3),  $\gamma$ -linolenic acid (18:3n-6), eicosapentaenoic acid (20:5n-3), arachidonic acid (20:4n-6), docosapentaenoic acid (22:5n-3) and docosatetraenoic acid (22:4n-6). Yeast cells were harvested, washed and dried, and fatty acid methyl esters (FAME) prepared by incubating the cells with 1 ml methylation reagent (10% v/v, 5% v/v 2,2-dimethylpropane and 85% v/v dry methanol) at 85 °C for 1 hour. The FAME were extracted, purified by thin layer chromatography (TLC), and analysed by gas chromatography (GC) all as described previously (Hastings et al., 2001). The proportion of substrate fatty acid converted to the longer chain fatty acid product was calculated from the gas chromatograms as  $100 \times [\text{product area}/(\text{product area} + \text{substrate area})]$ . Unequivocal confirmation of fatty acid products was performed by GC-mass spectrometry of the picolinyl derivatives as described in detail previously (Hastings et al., 2001).

## Materials

Eicosatetraenoic (20:4n-3), docosapentaenoic (22:5n-3) and docosatetraenoic (22:4n-6) acids (all > 98-99% pure) were purchased from Cayman Chemical Co., Ann Arbor, U.S.A. Tetracosapentaenoic (24:5n-3) and tetracosatetraenoic (24:4n-6) acids were both > 98% pure

provided by Dr. A.E.A. Porter, Department of Biological Sciences, University of Stirling and were chemically synthesised from 22:5n-3 and 22:4n-6, respectively, by successive C-1 additions, Linoleic (18:2n-6),  $\alpha$ -linolenic (18:3n-3),  $\gamma$ -linolenic (18:3n-6), stearidonic (18:4n-3), eicosatrienoic (20:3n-6), arachidonic (20:4n-6) and eicosapentaenoic (20:5n-3) acids (all >99% pure), butylated hydroxytoluene (BHT), 2,2-dimethoxypropane, galactose, nitrogen base, raffinose, tergitol NP-40 and uracil dropout medium were obtained from Sigma Chemical Co. Ltd., Dorset, UK. TLC (20 x 20 cm x 0.25 mm) plates precoated with silica gel 60 (without fluorescent indicator) were purchased from Merck, Darmstadt, Germany. All solvents were HPLC grade and were from Rathburn Chemicals, Peebleshire, U.K.

## RESULTS

### Salmon desaturase

Sequencing revealed that the salmon desaturase cDNA, SalDes, (Genbank accession number AF478472) included an open reading frame (ORF) of 1362 bp specifying a protein of 454 amino acids. The protein sequence possessed all the characteristic features of microsomal fatty acid desaturases, including three histidine boxes and two transmembrane regions (Fig. 1). The protein sequence also contained an N-terminal cytochrome  $b_5$  domain containing the haem-binding motif, H-P-G-G (alignment positions 63-66), similar to that of other fatty acid desaturases including the zebrafish desaturase. However, the salmon, as with the trout sequence, has an insertion of 10 amino acid residues at the N-terminal end. A pair-wise comparison was made between the identities and similarities of the fish and the human amino acid sequences (Table 1). The amino acid sequence predicted by the salmon ORF indicated that the desaturase candidate possessed 92% identity and 95% similarity to the amino acid sequence predicted by the trout Des cDNA (GenBank accession no. AF301910) and 64% identity and 80% similarity to the zebrafish  $\Delta 6/\Delta 5$  desaturase (GenBank accession no. AF309556). Comparisons between the salmon cDNA and the human  $\Delta 6$  (GenBank accession no. AF199596) and  $\Delta 5$  (GenBank accession no. AF126799) cDNAs, gave 60 and 63% identities with 75 and 78% similarities, respectively. Phylogenetic analysis comparing a variety of  $\Delta 5$  and  $\Delta 6$  desaturases clustered SalDes closely with other, as yet uncharacterised, salmonid genes (rainbow trout and cherry salmon) (Fig. 2). SalDes clustered slightly further from the zebrafish desaturase, which is known to possess both  $\Delta 6$  and  $\Delta 5$  activity. All the fish desaturase genes clustered together and closer to the mammalian (mouse and human)  $\Delta 6$  desaturases than to the  $\Delta 5$  desaturases (Fig. 2).

The salmon desaturase cDNA was functionally characterized by determining the fatty acid profiles of transformed *S. cerevisiae* containing either the pYES vector alone or the vector with the

SalDes insert (SalDes/pYES), grown in the presence of a variety of possible fatty acid substrates including 18:2n-6, 18:3n-3, 20:3n-6, 20:4n-3, 22:5n-3, 22:4n-6, 24:5n-3 and 24:4n-6. The fatty acid composition of the yeast transformed with the vector alone showed the four main fatty acids normally found in *S. cerevisiae*, namely 16:0, 16:1n-7, 18:0 and 18:1n-9, together with the exogenously derived substrate fatty acids, consistent with *S. cerevisiae* not possessing  $\Delta 5$  or  $\Delta 6$  fatty acid desaturase activities. Of all the fatty acid substrates tested, the most prominent additional peak was observed in the profile of SalDes/pYES-transformed yeast grown in the presence of the  $\Delta 5$  desaturase substrate, 20:4n-3 (Fig. 3B). Based on GC retention time, the additional peak associated with the presence of the salmon cDNA was identified as 20:5n-3. Approximately, 10% of 20:4n-3 was converted to 20:5n-3 in yeast transformed with the salmon desaturase (Table 2). Small amounts (< 1%) of 18:3n-3 (Fig.3A), 18:2n-6 and 20:3n-6 (Fig.3C) were also desaturated by the salmon clone. The identities of all product fatty acids were confirmed by GC-MS of the picolinyl derivatives. No desaturated products of C<sub>22</sub> fatty acid substrates were detected, indicating there was no  $\Delta 4$  desaturase activity (Table 2). Similarly, no traces of 24:6n-3 and 24:5n-6 were detected by GC-MS in yeast transformed with the salmon desaturase and grown in the presence of 24:5n-3 and 25:4n-6, respectively, indicating no  $\Delta 6$  activity towards C<sub>24</sub> substrates ( $\Delta 6^*$ ). These data indicate that the salmon enzyme is primarily an n-3  $\Delta 5$  desaturase with a low level of  $\Delta 6$  and n-6  $\Delta 5$  activity.

### Salmon elongase

Sequencing revealed that the salmon fatty acid elongase cDNA, SalElo, (Genbank accession number AY170327) included an open reading frame (ORF) of 888 bp specifying a protein of 295 amino acids. The protein sequence included characteristic features of microsomal fatty acid elongases, including a histidine box, ER retention signal and two transmembrane regions (Fig. 4). Comparison of the deduced amino acid sequence of the salmon elongase with that of elongases from zebrafish (ZfElo), mouse (Elov12) and human (ELOVL2 and ELOVL5) showed that SalElo shared 75, 55, 53 and 71 percent identity with ZfElo, Elov12, ELOVL2 and ELOVL5, respectively. Phylogenetic analyses of the salmon, zebrafish, human and mouse elongases showed the divergence from a common ancestor (Fig. 5). The fish elongases and human ELOV5 have diverged less from a common ancestor than have the two mammalian elongases ELOV2 and Elov2.

The salmon elongase cDNA was functionally characterized by determining the fatty acid profiles of transformed *S. cerevisiae* containing either empty vector alone or the vector with the salmon cDNA insert, grown in the presence of a variety of possible fatty acid substrates including 18:3n-6, 18:4n-3, 20:4n-6, 20:5n-3, 22:4n-6 and 22:5n-3. The results showed that the salmon elongase had very high C<sub>18-20</sub> elongase activity converting 71% of 18:4n-3 to 20:4n-3 (Fig.6A) and



42% of 18:3n-6 to 20:3n-6. The salmon clone also elongated 39% of 20:5n-3 (Fig. 6B) and 23% of 20:4n-6, thus demonstrating significant C<sub>20-22</sub> activity. The salmon clone was also able to elongate C<sub>22</sub> substrates such as 22:5n-3 and 22:4n-6 although at a much lower level than with C<sub>18</sub> and C<sub>20</sub> fatty acids (Fig.6C). These data indicate that the salmon elongase is a polyunsaturated fatty acid elongase active on a range of fatty acids with activity decreasing with chain length and with a preference for n-3 fatty acid substrates (Table 3). However, the salmon elongase also elongated monounsaturated fatty acids as evidenced by the presence of 18:1n-7, the elongation product of 16:1n-7, and small amounts of 20:1n-9 and 20:1n-7, the elongated products of the respective C<sub>18</sub> monoenes (Fig.6).

## DISCUSSION

The extent to which animals, including fish, can convert the plant-derived C<sub>18</sub> PUFA, 18:3n-3 and 18:2n-6, to long-chain C<sub>20/22</sub> HUFA varies with species and correlates with their complement of the microsomal fatty acyl desaturase and elongase enzymes. EPA is synthesised from 18:3n-3 by desaturation at the  $\Delta 6$  position, followed by a 2-carbon elongation, that is in turn followed by a further desaturation at the  $\Delta 5$  position (Cook, 1996). Synthesis of DHA from EPA has been suggested to proceed via a C<sub>24</sub> intermediate, requiring two successive elongations to 22:5n-3 and then 24:5n-3, which is then desaturated at the  $\Delta 6$  position (Sprecher et al., 1995). Here we report the sequences and functions of two cDNAs from Atlantic salmon that are involved in this fatty acid desaturation/elongation pathway. The salmon elongase cDNA (SalElo) encodes a protein that is very similar to mammalian elongases and has all the main structural characteristics possessed by elongases from other systems, notably the predicted transmembrane domains, the so-called histidine box (HXXHH) and the canonical C-terminal endoplasmic reticulum (ER) retention signal. Similarly, the protein sequence of the salmon desaturase (SalDes) has all the characteristics of microsomal fatty acid desaturases including three histidine boxes, a transmembrane region and the N-terminal cytochrome b<sub>5</sub> domain containing the haem-binding motif.

The salmon desaturase was cloned based on similarity to  $\Delta 6$  desaturases, and indeed it is slightly more similar at the protein sequence level to the human  $\Delta 6$  desaturase than to human  $\Delta 5$  desaturase. However, functional characterisation showed that the only significant activity that the desaturase had was towards 20:4n-3, producing 20:5n-3, and that it is, therefore, primarily a fatty acid  $\Delta 5$  desaturase. A small amount of  $\Delta 6$  desaturase activity was also detected but this is probably physiologically insignificant. The only other functionally characterised fish desaturase to date is that from zebrafish which was shown to be bifunctional having both  $\Delta 6$  and  $\Delta 5$  activities with  $\Delta 6 > \Delta 5$  (Hastings et al., 2001). The zebrafish gene was the first, and only, report of a fatty acid

desaturase in any species with both  $\Delta 5$  and  $\Delta 6$  activity and it was speculated that it may represent a component of a prototypic vertebrate PUFA biosynthesis pathway. The results presented here on salmon, a teleost fish more evolutionarily primitive than zebrafish, showing that fish can also have monofunctional desaturases indicates that the evolution of the HUFA biosynthetic pathway is possibly more complex. However, much more information will be required to elucidate this question and further desaturase cDNAs require to be isolated from fish species. It is probable that further desaturases remain to be isolated from Atlantic salmon. Indeed it is highly unlikely that the desaturase characterised in the present study, despite showing low levels of  $\Delta 6$  desaturase activity, is the only PUFA desaturase in salmon. In all our previous biochemical studies investigating fatty acid desaturation in salmon cells,  $\Delta 6$  activity was far greater than  $\Delta 5$  activity in both freshly isolated primary hepatocytes (Bell et al., 2001b, 2002; Tocher et al., 1997, 2000, 2001) and in established cell cultures (Tocher and Sargent, 1990). Therefore, Atlantic salmon should possess an enzyme with high  $\Delta 6$  desaturase activity, whether as a monofunctional enzyme or bifunctional like the previously characterised zebrafish gene. It was also acknowledged previously that other, monofunctional desaturases could be present in zebrafish (Hastings et al., 2001).

That salmon possess a  $\Delta 5$  desaturase is consistent with the fact that salmonids, including the anadromous Atlantic salmon that lives for much of its life in the marine environment, have all the enzymic activities required for the production of DHA. True marine fish are unable to produce DHA at a physiologically significant rate, and this has been attributed to the lack of  $\Delta 5$  desaturase activity in some species, such as sea bream (Tocher and Ghioni, 1999). In other marine species such as turbot, a deficiency in  $C_{18-20}$  elongation has been suggested (Ghioni et al., 1999), and clearly salmon also possess very active  $C_{18-20}$  fatty acid elongating activity. Indeed, this study has shown that all the fatty acid elongating activities,  $C_{18-20}$ ,  $C_{20-22}$  and  $C_{22-24}$ , required for the production of DHA are contained in one polypeptide in Atlantic salmon. In contrast to the desaturase activities, the fatty acid specificity of the salmon elongase is similar to that of the recently characterised zebrafish elongase (Agaba et al., 2003). Broad substrate specificity is a feature of vertebrate fatty acid elongases characterised to date. In contrast to the  $C_{18-20}$  elongases of *M. alpina* and *C. elegans* which show virtually no activity towards  $C_{20}$  PUFA (Parker-Barnes et al., 2000; Beaudoin et al., 2000), the human (ELOVL5) and rat elongases (rELO1) have high activity on 20:5n-3 and 20:4n-6 in addition to  $C_{18-20}$  elongase activity (Leonard et al., 2000b; Inagaki et al., 2002). However, the ELOVL5 and rELO1 elongases do not have the capacity to elongate  $C_{22}$  PUFA. More recently, two further mammalian genes have been cloned and characterized, a second human elongase (ELOVL2) and a mouse elongase (elovl2), both of which are able to elongate 22:5n-3 and 22:4n-6 to 24:5n-3 and 24:4n-6, respectively (Leonard et al., 2002). Like the salmon enzyme, the mouse elovl2 product is also able to elongate  $C_{18}$  and  $C_{20}$  PUFA in addition to  $C_{22}$  PUFA, whereas the human

ELOVL2 clone is only active towards C<sub>20</sub> and C<sub>22</sub> PUFA (Leonard et al., 2002). Thus, the salmon, zebrafish and mouse elovl2 elongases have similar PUFA specificities broader than the specificities of the human and rat elongases cloned so far. However, the presence of more than one PUFA elongase in human suggests that other elongases may yet be isolated from rat and mouse. Similarly, although the salmon elongase has a broader substrate specificity compared to either of the human genes that have overlapping substrate specificities, we cannot exclude the possibility that other fatty acid elongases will be isolated from salmon.

It is perhaps noteworthy that the salmon elongase could convert 22:5n-3 to 24:5n-3, as 24:5n-3 is an important intermediate in the biosynthesis of DHA in rats (Sprecher et al., 1995, 1999). Previously, biochemical studies had suggested that DHA synthesis in rainbow trout also proceeds via C<sub>24</sub> intermediates (Buzzi et al., 1996, 1997). The present study has revealed that a closely related salmonid, Atlantic salmon, possess a gene whose product is capable of producing C<sub>24</sub> PUFA from C<sub>22</sub> PUFA. Furthermore, direct synthesis of DHA from 22:5n-3 would require desaturation at the  $\Delta 4$  position, and thus a  $\Delta 4$  desaturase, as described in the marine microheterotroph, *Thraustochytrium sp* (Qui et al., 2001). However, desaturation of PUFA at the  $\Delta 4$  position has not been demonstrated in any vertebrate, and the desaturase gene isolated in the present study showed no  $\Delta 4$  desaturating activity.

Together with other published data, the results presented here prompt a question; do the differences between species in primary structures of desaturases and elongases contribute directly to the variation observed in the ability of the different species to biosynthesise HUFA? The protein sequences of the salmon  $\Delta 5$  desaturase, zebrafish bifunctional  $\Delta 6/\Delta 5$  desaturase, and the human  $\Delta 6$  and  $\Delta 5$  desaturases are all very similar, with large areas of conserved sequence. The catalytic activities are, of course, identical but the substrate specificities are quite different indicating that the differences in sequence have a significant impact. Similarly, the sequences of mammalian and fish elongases are similar, but those differences that occur clearly affect the substrate specificities of the enzymes. The accumulating sequence data presents an opportunity to study the structural features that have shaped PUFA desaturase and elongase substrate specificities during the evolution of vertebrates, and the possibility to artificially select for desaturases and elongases with superior specification. This may be supported by evidence from biochemical studies, which indicated that the inability to synthesise EPA and DHA in sea bream cells was due to very low  $\Delta 5$  desaturase activity (Tocher and Ghioni, 1999), whereas in turbot cells it was related to a deficiency in C<sub>18-20</sub> elongase (Ghioni et al., 1999), and that biosynthesis of arachidonic acid from 18:2n-6 in *M. alpina* may be rate-limited by the activity of the elongase (Wynne and Ratledge, 2000).

## ACKNOWLEDGEMENTS

This work was supported partly by a Scottish Higher Education Funding Council (SHEFC) Grant (“Aquagene”) and partly by the European Union Framework V Programme (project no. QLRT-2000-30058, “RAFOA”). NH was in receipt of a Natural Environment Research Council studentship and gratefully acknowledges the additional support of an award from the Scottish International Education Trust (SIET).

## REFERENCES

Agaba, M., Tocher, D.R., Dickson, C., Dick, J.R., and Teale, A.J. (2003). A zebrafish cDNA encoding a multifunctional enzyme involved in the elongation of polyunsaturated, monounsaturated and saturated fatty acids. *Mar Biotechnol* in press.

Aki, T., Shimada, Y., Inagaki, K., Higashimoto, H., Kawamoto, S., Shiget, S., Ono, K., and Suzuki, O. (1999). Molecular cloning and functional characterisation of rat  $\Delta 6$  fatty acid desaturase. *Biochem Biophys Res Commun* 255: 575-579.

Barlow, S. (2000). Fishmeal and fish oil: sustainable feed ingredients for aquafeeds. *Global Aquacult Advocate* 4: 85-88.

Beaudoin, F., Michaelson, L.V., Hey, S.J., Lewis M.L., Shewry, P.R., Sayanova, O., and Napier, J.A. (2000). Heterologous expression in yeast of the polyunsaturated fatty acid biosynthetic pathway. *Proc Natl Acad Sci U S A* 97: 6421-6426.

Bell, J.G., McEvoy, J., Tocher, D.R., McGhee, F., Campbell, P.J. and Sargent, J.R. (2001b). Replacement of fish oil with rape seed oil in diets of Atlantic salmon (*Salmo salar*) affects tissue lipid compositions and hepatocyte fatty acid metabolism. *J Nutr* 131: 1535-1543.

Bell, J.G., Henderson, R.J., Tocher, D.R., McGhee, F., Dick, J.R., Porter, A., Smullen, R. and Sargent, J.R. (2002). Substituting fish oil with crude palm oil in the diet of Atlantic salmon (*Salmo salar*) affects tissue fatty acid compositions and hepatic fatty acid metabolism. *J Nutr* 132: 222-230.

Bell, J.G., Tocher, D.R., Farndale, B.M., Cox, D.I., McKinney, R.W. and Sargent, J.R. (1997). The effect of dietary lipid on polyunsaturated fatty acid metabolism in Atlantic salmon (*Salmo salar*) undergoing parr-smolt transformation. *Lipids* 32: 515-525.

- Bell, M.V., Dick, J.R. and Porter, A.E.A. (2001a). Biosynthesis and tissue deposition of docosahexaenoic acid (22:6n-3) in rainbow trout (*Oncorhynchus mykiss*). *Lipids* 36: 1153-1159.
- Buzzi, M., Henderson, R.J. and Sargent, J.R. (1996). The desaturation and elongation of linolenic acid and eicosapentaenoic acid by hepatocytes and liver microsomes from rainbow trout (*Oncorhynchus mykiss*) fed diets containing fish oil or olive oil. *Biochim Biophys Acta* 1299: 235-244.
- Buzzi, M., Henderson, R.J. and Sargent, J.R. (1997). Biosynthesis of docosahexaenoic acid in trout hepatocytes proceeds via 24-carbon intermediates. *Comp Biochem Physiol* 116B: 263-267.
- Cho, H.P., Nakamura, M.T. and Clarke, S.D. (1999a). Cloning expression and nutritional regulation of the human  $\Delta 6$  desaturase. *J Biol Chem* 274: 471-477.
- Cho, H.P., Nakamura, M.T. and Clarke, S.D. (1999b). Cloning expression and nutritional regulation of the human  $\Delta 5$  desaturase. *J Biol Chem* 274: 37335-37339.
- Cook, H.W. (1996). Fatty acid desaturation and chain elongation in eukaryote. In: *Biochemistry of Lipids, Lipoproteins and Membranes*, Vance, D.E and Vance, J.E. (eds). Amsterdam: Elsevier, 129-152.
- Ghioni, C., Tocher, D.R., Bell, M.V., Dick, J.R. and Sargent, J.R. (1999). Low C<sub>18</sub> to C<sub>20</sub> fatty acid elongase activity and limited conversion of stearidonic acid, 18:4n-3, eicosapentaenoic acid, 20:5n-3, in a cell line from the turbot, *Scophthalmus maximus*. *Biochim Biophys Acta* 1437: 170-181.
- Hastings, N., Agaba, M., Tocher, D.R., Leaver, M.J., Dick, J.R., Sargent, J.R. and Teale, A.J. (2001). A vertebrate fatty acid desaturase with  $\Delta 5$  and  $\Delta 6$  activities. *Proc Natl Acad Sci U S A* 98: 14304-14309.
- Huang, Y.-S., Chaudhary, S., Thurmond, J., Bobik, E.G., Yuan, L., Chan, G.E., Kirchner, S.J., Mukerji, P. and Knutson, D.S. (1999). Cloning of  $\Delta 12$ - and  $\Delta 5$ -desaturases from *Mortierella alpina* and recombinant production of  $\gamma$ -linolenic acid in *Saccharomyces cerevisiae*. *Lipids* 34: 649-659.

Inagaki, K., Aki, T., Fukuda, Y., Kawamoto, S., Shigeta, S., Ono, K. and Suzuki, O. (2002). Identification and expression of a rat fatty acid elongase involved the biosynthesis of C18 fatty acids. *Biosci Biotechnol Biochem* 66: 613-621.

Kyte, J. and Doolittle, R. (1982) A simple method for displaying the hydropathic character of a protein. *J. Mol. Biol.* 157: 105-132.

Leonard, A.E., Kelder, B., Bobik, E.G., Kroeger, P.E., Chuang, L.-T., Thurmond, J.M., Parker-Barnes, J.M., Kopchick, J.J., Huang, Y.-S. and Murkerji, P. (2000a). cDNA cloning and characterisation of human  $\Delta 5$  desaturase involved in the synthesis of arachidonic acid. *Biochem J* 347: 719-724.

Leonard, A.E., Bobik, E.G., Dorado, J., Kroeger, P.E., Chuang, L.-T., Thurmond, J.M., Parker-Barnes, J.M., Das, T., Huang Y.-S. and Murkerji, P. (2000b). Cloning of a human cDNA encoding a novel enzyme involved in the elongation of long chain polyunsaturated fatty acids. *Biochem J* 350: 765-770.

Leonard, A.E., Kelder, B., Bobik, E.G., Chuang, L.-T., Lewis, C.J., Kopchick, J.J., Murkerji, P. and Huang, Y.-S. (2002). Identification and expression of mammalian long-chain PUFA elongation enzymes. *Lipids* 37: 733-740.

Michaelson, L.V., Lazarus, C.M., Griffiths, G., Napier, J.A. and Stobart, A.K. (1998a). Isolation of a  $\Delta 5$  fatty acid desaturase gene from *Mortierella alpina*. *J Biol Chem* 273: 19055-19059.

Michaelson, L.V., Napier, J.A., Lewis, M., Griffiths, G., Lazarus, C.M. and Stobart, A.K. (1998b). Functional identification of a fatty acid  $\Delta^5$  desaturase gene from *Caenorhabditis elegans*. *FEBS Lett* 439: 215-218.

Napier, J.A., Hey, S.J., Lacey, D.J. and Shewry, P.R. (1998). Identification of a *Caenorhabditis elegans*  $\Delta 6$  fatty acid - desaturase by heterologous expression in *Saccharomyces cerevisiae*. *Biochem J* 330: 611-614.

Parker-Barnes, J.M., Das, T., Bobik, E., Leonard, A.E., Thurmond, J.M., Chuang, L., Huang, Y.S. and Mukerji, P. (2000). Identification and characterization of an enzyme involved in the elongation of n-6 and n-3 polyunsaturated fatty acid. *Proc Natl Acad Sci USA* 97: 8284-8289.

Qui, X., Hong, H. and MacKenzie, S.L. (2001). Identification of a  $\Delta 4$  fatty acid desaturase from *Thraustochytrium* sp. involved in the synthesis of docosahexaenoic acid by heterologous expression in *Saccharomyces cerevisiae* and *Brassica juncea*. *J Biol Chem* 276: 31561-31566.

Saitou, N. and Nei, M. (1987). The neighbor-joining method. A new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4: 406-425.

Sargent, J.R. and Tacon, A. (1999). Development of farmed fish: a nutritionally necessary alternative to meat. *Proc Nutr Soc* 58: 377-383.

Sargent, J.R., Henderson, R.J. and Tocher, D.R. (1989). The lipids. In: *Fish Nutrition*. Second edition, Halver, J.E., (ed). San Diego:Academic Press, 153-218.

Sargent, J.R., Tocher, D.R. and Bell, J.G. (2002). The lipids. In: *Fish Nutrition*. Third edition, Halver, J. E. and Hardy, R.W. (eds). San Diego:Academic Press, 181-257.

Sprecher, H. and Chen, Q. (1999). Polyunsaturated fatty acid biosynthesis: a microsomal - peroxisomal process. *Prostaglandins Leukotrienes Essent Fatty Acids* 60: 317-321.

Sprecher, H., Luthria, D.L., Mohammed, B.S. and Baykousheva, S.P. (1995). Re-evaluation of the pathways for the biosynthesis of polyunsaturated fatty acids. *J Lipid Res* 36: 2471 -2477.

Tidwell, J.H. and Allan, G.L. (2002). Fish as food: aquaculture's contribution. *World Aquaculture* 33: 44-48.

Tocher, D.R. and Ghioni, C. (1999). Fatty acid metabolism in marine fish: Low activity of  $\Delta 5$  desaturation in gilthead sea bream (*Sparus aurata*) cells. *Lipids* 34: 433-440.

Tocher, D.R. and Sargent, J.R. (1990). Incorporation into phospholipid classes and metabolism via desaturation and elongation of various  $^{14}\text{C}$ -labelled (n-3) and (n 6) polyunsaturated fatty acids in trout astrocytes in primary culture. *J Neurochem* 54: 2118-2124.

Tocher, D.R., Bell, J.G., Dick, J.R. and Sargent, J.R. (1997). Fatty acyl desaturation in isolated hepatocytes from Atlantic salmon (*Salmo salar*): Stimulation by dietary borage oil containing  $\gamma$ -

linolenic acid. *Lipids* 32: 1237-1247.

Tocher, D.R., Bell, J.G., Henderson, R.J., McGhee, F., Mitchell, D. and Morris, P.C. (2000). The effect of dietary linseed and rapeseed oils on polyunsaturated fatty acid metabolism in Atlantic salmon (*Salmo salar*) undergoing parr-smolt transformation. *Fish Physiol Biochem* 23: 59-73.

Tocher, D.R., Bell, J.G., MacGlaughlin, P., McGhee, F. and Dick, J.R. (2001). Hepatocyte fatty acid desaturation and polyunsaturated fatty acid composition of liver in salmonids: Effects of dietary vegetable oil. *Comp Biochem Physiol* 130: 257-270.

Tocher, D.R., Leaver, M.J. and Hodgson, P.A. (1998). Recent advances in the biochemistry and molecular biology of fatty acyl desaturases. *Prog Lipid Res* 37: 73-117.

Watts, J.L. and Browse, J. (1999). Isolation and characterisation of a  $\Delta 5$  fatty acid desaturase from *Caenorhabditis elegans*. *Arch Biochem Biophys* 362: 175-182.

Wilson, R.P. (1989). Amino acids and proteins. In: *Fish Nutrition*. second edition, Halver, J.E. (ed). San Diego:Academic Press, 153-218.

Wynn, J.P. and Ratledge, C. (2000). Evidence that the rate limiting step for the biosynthesis of arachidonic acid in *Mortierella alpina* is at the level of the 18:3 to 20:3 elongase. *Microbiology* 146: 2325-2331.



## Legends to Figures

Fig.1. Comparison of the deduced amino acid sequence of a polyunsaturated fatty acyl desaturase from Atlantic salmon with that of desaturases from trout (*O. mykiss*), zebrafish (*Danio rerio*) and human. Identical residues are shaded black and similar residues are shaded grey. Identity/similarity shading was based on the BLOSUM62 matrix and the cut off for shading was 75%. Characteristic features are denoted thus, histidine boxes (underline), cytochrome b<sub>5</sub>-domain (dotted underline with asterisks denoting the haem-binding motif) and transmembrane regions (dashed underline). Transmembrane regions were predicted from a hydropathy plot taking peaks with scores > 1.6 using a scan window size of 18 (Kyte and Doolittle, 1982).

Fig.2. Phylogenetic tree of desaturases from salmon, other fish species (zebrafish, trout, carp, cherry salmon and tilapia), mammals (mouse and human), yeast (*Mortierella alpina*) and nematode (*Caenorhabditis elegans*). The tree was constructed using the N-J method using *CLUSTALX* and *NJPLLOT*. The horizontal branch length is proportional amino acid substitution rate per site. The numbers represent the frequencies with which the tree topology presented here was replicated after 1000 bootstrap iterations. An asterix denotes a desaturase sequence that has not yet been functionally characterized.

Fig.3. Identification of fatty acid desaturation products in transgenic yeast (*Saccharomyces cerevisiae*). Fatty acids were extracted from yeast transformed with SalDes/pYES grown in the presence of either A: 18:3n-3, B: 20:4n-3 or C: 20:3n-6. The first four peaks in panels A-C are the main endogenous fatty acids of *S. cerevisiae*, namely 16:0 (1), 16:1n-7 (2), 18:0 (3) and 18:1n-9 (4). The additional peaks in each panel are the exogenously added substrate fatty acids and the resultant desaturated products, namely 18:3n-3 (5), 18:4n-3 (6), 20:4n-3 (7), 20:5n-3 (8), 20:3n-6 (9) and 20:4n-6 (10).

Fig.4. Comparison of the deduced amino acid sequence of a polyunsaturated fatty acyl elongase from Atlantic salmon elongase (SalElo) with that of elongases from zebrafish (ZfElo), mouse (Elov12) and human (ELOVL2 and ELOVL5). Identical residues are shaded black and similar residues are shaded grey. Identity/similarity shading was based on the BLOSUM62 matrix and the cut off for shading was 75%. Characteristic features are denoted thus, histidine boxes (underline), ER retention signal (asterisks) and transmembrane regions (dashed underline). Transmembrane regions were predicted from a hydropathy plot taking peaks with scores > 1.6 using a scan window size of 18 (Kyte and Doolittle, 1982).

Fig.5. Phylogenetic tree of elongases from salmon (SalElo), zebrafish (ZfElo), human (ELOVL2, ELOVL2) and mouse (Elovl2). The tree was constructed using the N-J method using *CLUSTALX* and *NJPLLOT*. The horizontal branch length is proportional amino acid substitution rate per site. The numbers represent the percentage frequencies with which the tree topology presented here was replicated after 10000 bootstrap iterations.

Fig.6. Identification of fatty acid elongation products in transgenic yeast (*Saccharomyces cerevisiae*). Fatty acids were extracted from yeast transformed with SalElo/pYX222 grown in the presence of either A: 18:4n-3, B: 20:5n-3 or C: 22:5n-3. The first four peaks in panels A-C are the main endogenous fatty acids of *S. cerevisiae*, namely 16:0 (1), 16:1n-7 (2), 18:0 (3) and 18:1n-9 (4). The additional peaks in each panel are the exogenously added substrate fatty acids and the resultant elongated products, namely 18:4n-3 (5), 20:4n-3 (6), 20:5n-3 (7), 22:5n-3 (8) and 24:5n-3 (9).

Fig.1.

Atlantic salmon D5	MCGCGQQTESSEPAKCDGLEPDGCGGCSAVYTWEVQPHSHRSDQWLVIDRKVYNTITQAKRHPGCGIRVISHFACEDATE	80
Rainbow trout D6	MCGCGQQTESSEPAKCDGVCPDGGGCGGSAVYTWEVQPHCHRSQWLVIDRKVYNTITQAKRHPGCGIRVISHFACEDATE	80
Zebrafish D5/D6	MCGCGQQTDRTDITNC-----RFSSTWEEVQPHHKHCDQWVVERKVYNTSQVVERHPGCGIRLICHYACEDATE	70
Human D6 (Fads2)	MCRGCMQCECAAREVS-----VPTFSWEEVQPHMLRTDQWLVIDRKVYNTITQMSIQHPGCGIRVICHYACEDATE	70
Human D5 (Fads1)	MADPLAAETAAQCLTP-----RYFTQDEVAQRSGCEERQWLVIDRKVYNTISEITPRHPGCGSEVISHYACEDATE	69
*****		
Atlantic salmon D5	AFSAFHLDANFVVKFLKPLLCGLAPTEPSQDHGKRMALWDFQALRDHYEREGLLRARLLFFSLYLCHILLLEALALGL	160
Rainbow trout D6	AFVAFHPDPNFVVKFLKPLLCGLAPTEPSQDHGKRMALWDFQALRDHYEREGLLRARPLFFSLYLCHILLLEALALGL	160
Zebrafish D5/D6	AFTAFHPNLQLVKKYKPLLCGLLEASPSQDRQKRMALWDFQALRERLEAECCFKTQPLFFALELCHILLLEALAFMM	150
Human D6 (Fads2)	AFRAFHPDLEFVCKFLKPLLCGLAPTEPSQDHGKRMALWDFQALRERLEAECCFKTQPLFFALELCHILLLEALAFMM	150
Human D5 (Fads1)	PEVAFHINKRLVKKYKPLLCGLLEASPSQDHGKRMALWDFQALRERLEAECCFKTQPLFFALELCHILLLEALAFMM	149
*****		
Atlantic salmon D5	LWVVGTSQSLTLLCSLMLATSQAQAGWLQHDVCHLSVCRTSSQNHVLDHKFVIGHLKGCASAGWNNHRRFQHHAKPMVFREK	240
Rainbow trout D6	LWVVGTSQSLTLLCSLMLATSQAQAGWLQHDVCHLSVCRTSSQNHVLDHKFVIGHLKGCASAGWNNHRRFQHHAKPMVFREK	240
Zebrafish D5/D6	WVYFGTGQINTLIVAVILATAQAQAGWLQHDVCHLSVVFRTSGMNHLDHKFVIGHLKGCASAGWNNHRRFQHHAKPMVFREK	230
Human D6 (Fads2)	VFYFGNGQIPIITLITAFVLATSQAQAGWLQHDVCHLSVYRKPMNHLDHKFVIGHLKGCASAGWNNHRRFQHHAKPMVFREK	230
Human D5 (Fads1)	LWVVGTSQSLPFLCAVLLSAVQAQAGWLQHDVCHLSVVFRTSGMNHLDHKFVIGHLKGCASAGWNNHRRFQHHAKPMVFREK	229
*****		
Atlantic salmon D5	PDVNSLP-WFVLGDTQPVVEYGIKKKPYHQQHYFFLICPPLIPVVFETIQIRTFMFSQRDWDVLAWSMSFYDRFFCC	319
Rainbow trout D6	PDVNSLP-WFVLGDTQPVVEYGIKKKPYHQQHYFFLICPPLIPVVFETIQIRTFMFSQRDWDVLAWSMSFYDRFFCC	319
Zebrafish D5/D6	PDVNSLP-WFVLGDTQPVVEYGIKKKPYHQQHYFFLICPPLIPVVFETIQIRTFMFSQRDWDVLAWSMSFYDRFFCC	309
Human D6 (Fads2)	PDVNSLP-WFVLGDTQPVVEYGIKKKPYHQQHYFFLICPPLIPVVFETIQIRTFMFSQRDWDVLAWSMSFYDRFFCC	309
Human D5 (Fads1)	PDVNSLP-WFVLGDTQPVVEYGIKKKPYHQQHYFFLICPPLIPVVFETIQIRTFMFSQRDWDVLAWSMSFYDRFFCC	309
*****		
Atlantic salmon D5	YYPFGCFGCSVALISFVRFLSHWVFWVTQMNHLPMEIDHERHODMLTMQLSATCNIEQSTFNDWFSCHLNFQIEHHLLFP	399
Rainbow trout D6	YYPFGCFGCSVALISFVRFLSHWVFWVTQMNHLPMEIDHERHODMLTMQLSATCNIEQSTFNDWFSCHLNFQIEHHLLFP	399
Zebrafish D5/D6	YYPFGCFGCSVALISFVRFLSHWVFWVTQMNHLPMEIDHERHODMLTMQLSATCNIEQSTFNDWFSCHLNFQIEHHLLFP	389
Human D6 (Fads2)	YYPFGCFGCSVALISFVRFLSHWVFWVTQMNHLPMEIDHERHODMLTMQLSATCNIEQSTFNDWFSCHLNFQIEHHLLFP	389
Human D5 (Fads1)	YYPFGCFGCSVALISFVRFLSHWVFWVTQMNHLPMEIDHERHODMLTMQLSATCNIEQSTFNDWFSCHLNFQIEHHLLFP	389
*****		
Atlantic salmon D5	TMPRHNYELVAPLVRLCEKHGVPYQVETLQKCMTDVVRSLRKSGLWLDAYLHK	454
Rainbow trout D6	TMPRHNYELVAPLVRLCEKHGVPYQVETLQKCMTDVVRSLRKSGLWLDAYLHK	454
Zebrafish D5/D6	TMPRHNYELVAPLVRLCEKHGVPYQVETLQKCMTDVVRSLRKSGLWLDAYLHK	444
Human D6 (Fads2)	TMPRHNYELVAPLVRLCEKHGVPYQVETLQKCMTDVVRSLRKSGLWLDAYLHK	444
Human D5 (Fads1)	TMPRHNYELVAPLVRLCEKHGVPYQVETLQKCMTDVVRSLRKSGLWLDAYLHK	444

**Fig. 2**

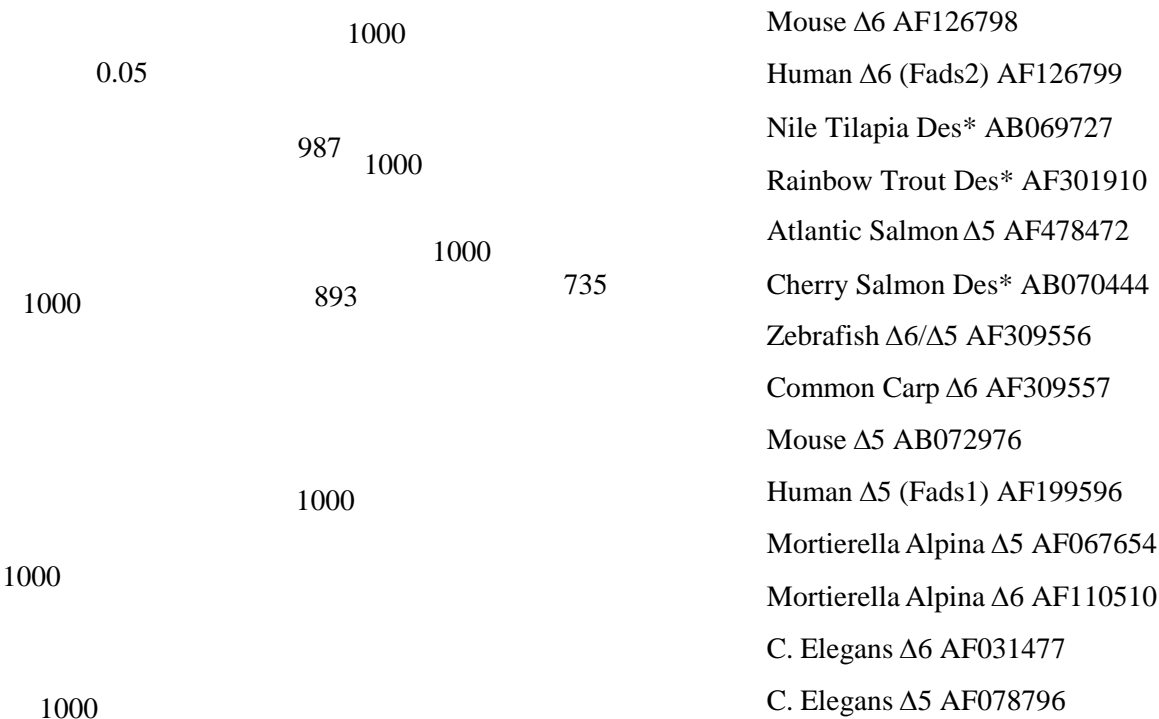
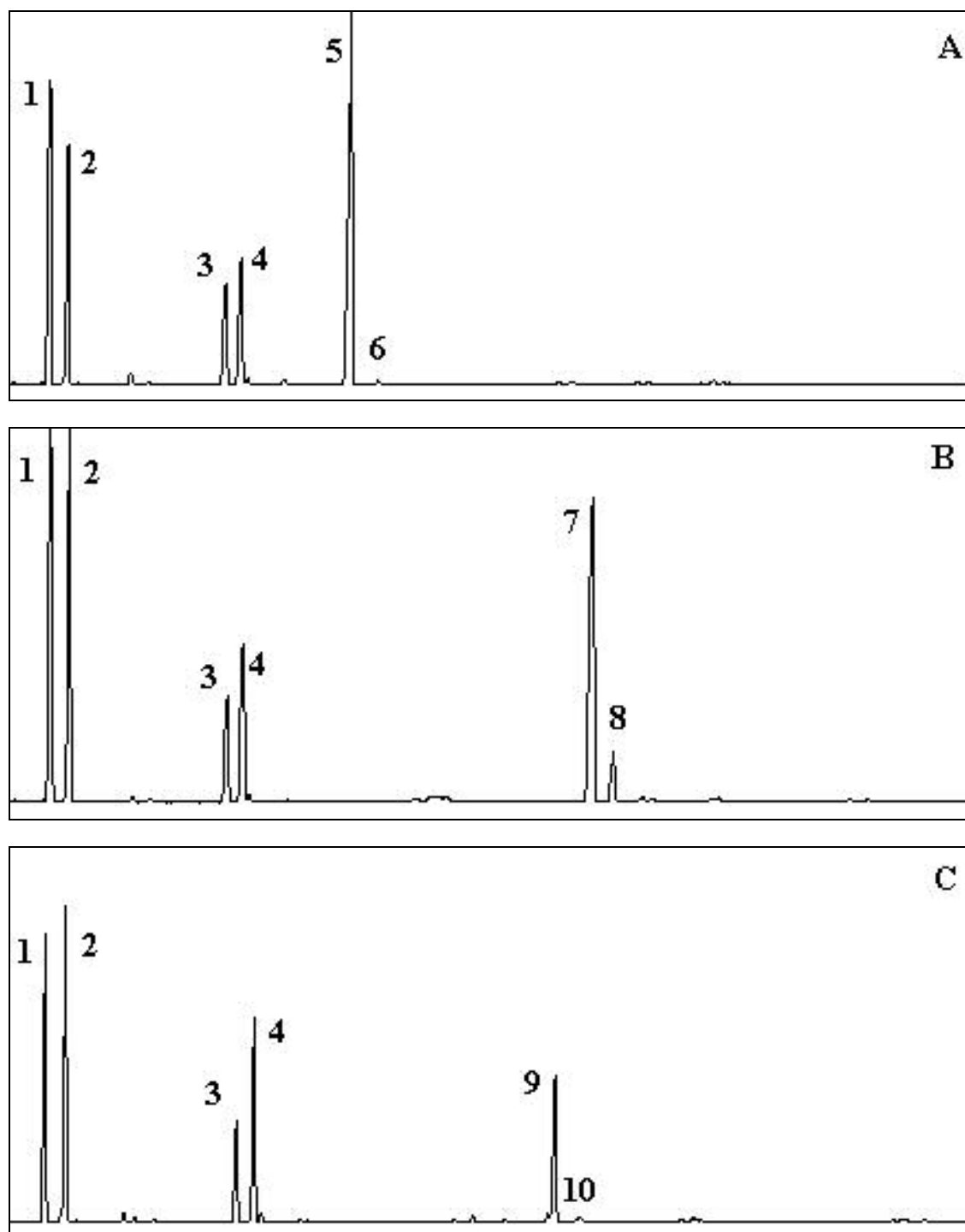


Fig.3



**Fig.4**



**Fig.5.**

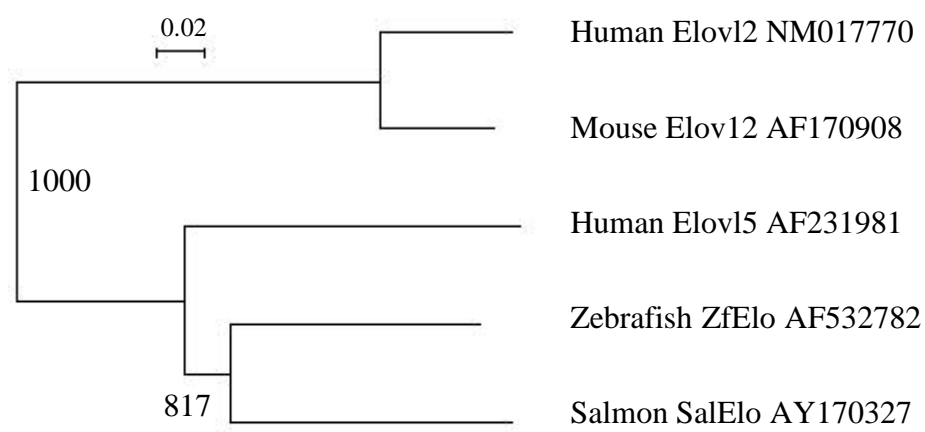


Fig.6.

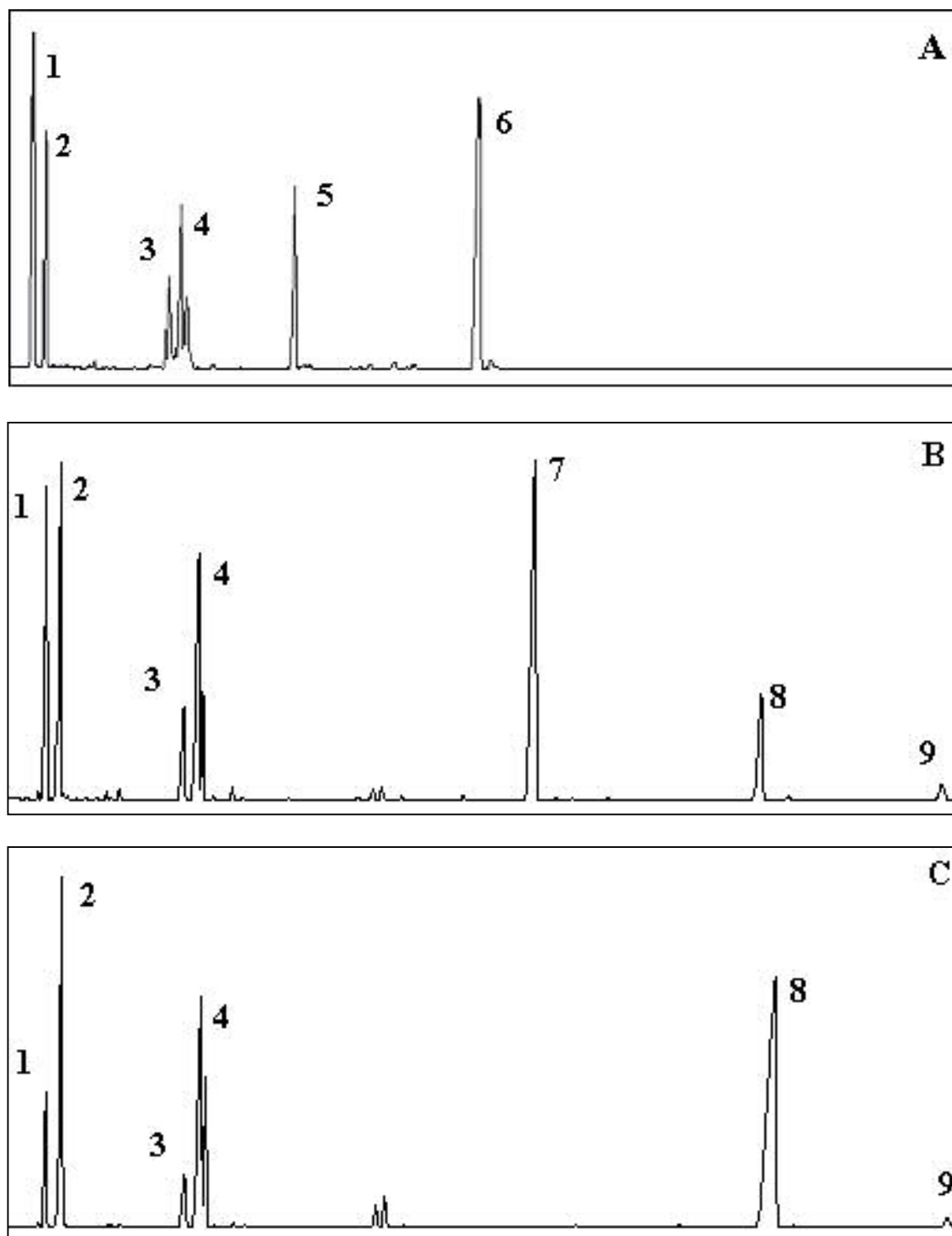




Table 1. Pair-wise comparison between the identities and similarities of the amino acid sequences of fish and human fatty acid desaturases.

	Human $\Delta 5$	Salmon (SalDes)	Trout Des	Zebrafish $\Delta 6/\Delta 5$
Human $\Delta 6$	62 (76)	60 (75)	61 (76)	58 (75)
Human $\Delta 5$		63 (78)	65 (79)	64 (78)
Salmon (SalDes)			92 (95)	64 (80)
Trout Des				66 (81)

Data are percentages of amino acid residues that are identical or similar (in brackets). Similar amino acid residues are defined as having the same physical/chemical characteristics (e.g. basic, acidic, hydrophobic, hydrophilic). Trout Des, putative  $\Delta 6$  desaturase from rainbow trout.

Table 2. Functional characterisation of Atlantic salmon desaturase (SalDes) in *Saccharomyces cerevisiae* .

Fatty acid substrate	Product	Conversion (%)	Activity
18:3n-3	18:4n-3	0.6	$\Delta 6$
18:2n-6	18:3n-6	0.4	$\Delta 6$
20:4n-3	20:5n-3	10.2	$\Delta 5$
20:3n-6	20:4n-6	0.9	$\Delta 5$
22:5n-3	22:6n-3	n.d.	$\Delta 4$
22:4n-6	22:5n-6	n.d.	$\Delta 4$
24:5n-3	24:6n-3	n.d.	$\Delta 6^*$
24:4n-6	24:5n-6	n.d.	$\Delta 6^*$

Results are expressed as a percentage of substrate fatty acid converted to desaturated product.  $\Delta 6^*$ ,  $\Delta 6$  desaturase activity towards C<sub>24</sub> fatty acid substrates. n.d., not detected.

Table 3. Functional characterisation of Atlantic salmon elongase (SalElo) in *Saccharomyces cerevisiae* .

Fatty acid substrate	Product	Conversion (%)	Activity
18:4n-3	20:4n-3	71.8	C <sub>18-20</sub>
18:3n-6	20:3n-6	42.1	C <sub>18-20</sub>
20:5n-3	22:5n-3	38.9	C <sub>20-22</sub>
20:4n-6	22:4n-6	22.6	C <sub>20-22</sub>
22:5n-3	24:6n-3	0.7 (2.9)	C <sub>22-24</sub>
22:4n-6	24:5n-6	0.3 (2.0)	C <sub>22-24</sub>

Results are expressed as a percentage of substrate fatty acid converted to elongated product. Two values are shown for conversion of the C<sub>22</sub> fatty acid substrates, with the first values being the percentage conversion when the C<sub>22</sub> substrates themselves were used, whereas the values in brackets represent the percentage conversion of C<sub>22</sub> produced when the yeast were grown in C<sub>20</sub> substrates.